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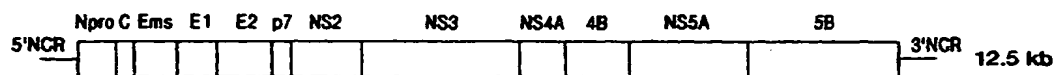
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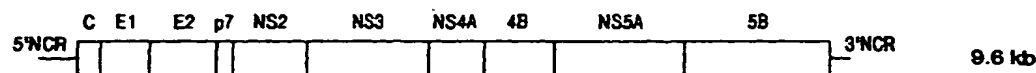
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(54) Title: HCV/BVDV CHIMERIC GENOMES AND USES THEREOF

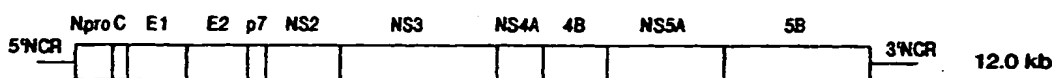
BVDV-NADL



HCV-H77C



HCV/BVDV (Chimeric RNA)



(57) Abstract: The present invention relates to molecular approaches to the production of nucleic acid sequences which comprise the genomes of chimeric hepatitis C virus-bovine viral diarrhea viruses (HCV-BVDV). The invention also relates to the use of these chimeric nucleic acid sequences to produce chimeric virions in cells and the use of these chimeric virions in HCV antibody neutralization assays, and for the development of vaccines and therapeutics for HCV.

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TITLE OF INVENTION

HCV/BVDV Chimeric Genomes and Uses Thereof

FIELD OF INVENTION

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The present invention relates to molecular approaches to the production of nucleic acid sequences which comprise the genomes of chimeric hepatitis C virus-bovine viral diarrhea viruses (HCV-BVDV). The invention also relates to the use of these chimeric nucleic acid sequences to produce chimeric virions in cells and the use of these chimeric virions in HCV antibody neutralization assays, and for the development of vaccines and therapeutics for HCV.

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Background Of Invention

Hepatitis C virus (HCV) has a positive-sense single-strand RNA genome and is a member of the genus Hepacivirus within the Flaviviridae family of viruses (Rice, 1996). As for all positive-stranded RNA viruses, the genome of HCV functions as mRNA from which all viral proteins necessary for propagation are translated.

The viral genome of HCV is approximately 9600 nucleotides (nts) in length and consists of a highly conserved 5' untranslated region (UTR), a single long open reading frame (ORF) of approximately 9,000 nts and a complex 3' UTR. The 5' UTR contains an internal ribosomal entry site (Tsukiyama-Kohara et al., 1992; Honda et al., 1996). The 3' UTR consists of a short variable region, a polypyrimidine tract of variable length and, at the 3' end, a highly conserved region of approximately 100 nucleotides (Kolykhalov et al., 1996;

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accounts for the majority of HCV infections but genotypes 2 and 3 each account for 5-15%.

At present, more than 80% of individuals infected with HCV become chronically infected and these chronically infected individuals have a relatively high risk of developing chronic hepatitis, liver cirrhosis and hepatocellular carcinoma (Hoofnagle, 1997). The only effective therapy for chronic hepatitis C, interferon (IFN), alone or in combination with ribavirin, induces a sustained response in less than 50% of treated patients (Davis et al., 1998; McHutchinson et al., 1998). Consequently, HCV is currently the most common cause of end stage liver failure and the reason for about 30% of liver transplants performed in the U.S. (Hoofnagle, 1997). In addition, a number of recent studies suggested that the severity of liver disease and the outcome of therapy may be genotype-dependent (reviewed in Bukh et al., 1997). In particular, these studies suggested that infection with HCV genotype 1b was associated with more severe liver disease (Brecht, 1997) and a poorer response to IFN therapy (Fried and Hoofnagle, 1995). As a result of the inability to develop a universally effective therapy against HCV infection, it is estimated that there are still more than 25,000 new infections yearly in the U.S. (Alter 1997) Moreover, since there is no vaccine for HCV, HCV remains a serious public health problem.

Despite the intense interest in the development of vaccines and therapies for HCV, progress has been hindered by the absence of a useful cell culture system for laboratory study (2-7). For example, although the virus has been grown in some cell lines,

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chimeric nucleic acid sequences to study the molecular properties of HCV indirectly in vitro.

The present invention also relates to the polypeptides encoded by the chimeric nucleic acid sequences of the invention or fragments thereof.

The invention also provides that the chimeric nucleic acid sequences and the chimeric viruses of the invention be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

DESCRIPTION OF FIGURES

Fig. 1. Genomic organization of BVDV, HCV and HCV/BVDV chimera. The BVDV and HCV are NADL (14, 21) and H77 strains (12), respectively. The complete BVDV-NADL genome consists of, in 5' to 3' order, 5'NCR (nucleotides 1-385), N^{Pro} (nucleotides 386-889), Core (nucleotides 890-1195), E^{ns} (nucleotides 1196-1876), E1 (nucleotides 1877-2461), E2 (nucleotides 2462-3583), P7 and nonstructural genes (nucleotides 3584-12349) and 3'NCR (nucleotides 12352-12578).

Fig. 2. Strategy for the construction of chimeric cDNA, pHCV/BVDV-3, which has core, E1 and E2 of HCV in the backbone of BVDV. The fusion PCR products were cloned into pBV18-F2 after digestion with *Sna*B I and *Bsm* I. The fragments containing fusion PCR products were cloned into pSDMlu-3' after digestion with *Cla* I and *Dra* III.

Figures 3A-3H show the nucleotide and deduced amino acid sequences of the infectious HCV clone of genotype 1a.

Figures 4A-4H show the nucleotide and deduced amino acid sequences of the infectious clone of genotype

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constructed. Such chimeras can be used to determine the relative importance of E1 or E2 for infection of cell lines. In another embodiment, HCV/BVDV chimeras in which one of the nonstructural genes of BVDV, such as NS3 RNA helicase, NS3 protease, or the NS5B RNA-dependent RNA polymerase are replaced by the corresponding non-structural genes of HCV may be constructed. Such chimeras would, for example, be useful in identifying inhibitors of viral enzyme activity which would be useful as antiviral agents.

In yet another embodiment, hypervariable region 1 (HVR1) from multiple HCV genotypes may be combined into one HCV/BVDV chimera. The only limit for constructing this type of chimera is that the viral genome must be able to be packaged. Alternatively, a chimera can be constructed which contain an HVR1 sequence from one HCV genotype. Such chimeras can be used as an inactivated multivalent vaccine or to screen for neutralizing antibodies to multiple HCV genotypes.

The HCV/BVDV chimeras of the invention may be constructed using any HCV and BVDV clones. However, in a preferred embodiment, the HCV clones are infectious HCV clones of genotype 1a (ATCC accession number PTA-157; Figures 3A-3F), 1b (ATCC accession number 209596; Figures 4A-4F) or 2a (ATCC accession number PTA-153; SEQ ID NOS:3-4) and the infectious BVDV clone pVVNADL are used.

In constructing the chimeric nucleic acid sequences of the invention, it is to be understood that the retention of the E^{ns} gene of BVDV in any chimeric is entirely optional. Thus, when it is stated that the HCV/BVDV chimeras could be constructed in which, for

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the growing of animal cells in vitro and transfecting the cells with the chimeric nucleic acid of the invention, then determining if the cells show indicia of HCV infection. Such indicia include the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the detection of viral polypeptides by Western blotting using antibodies specific therefor; and the detection of newly transcribed viral RNA within the cells via methods such as RT-PCR. The presence of live, infectious virus particles following such tests may also be shown by injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of HCV infection. Alternatively, the presence of live, infectious virus particles following such tests may also be shown by serial passaging the chimeric virus in cells.

Suitable cells or cell lines for culturing the chimeric viruses of the invention include, but are not limited to, EBTr(A) and Huh7.

Preferably, transfection of cells with the chimeric sequences is carried out in the presence of helper BVDV which is preferably of a noncytopathogenic strain. In one embodiment, the cell lines to be infected may already contain a helper BVDV. Such cells include, but are not limited to, EBTr(A).

Alternatively, the cell lines to be transfected may be infected with a helper BVDV prior to, or concurrent with, transfection with the chimeric sequences of the invention.

The present invention also relates to polypeptides encoded by the chimeric nucleic acid

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can then be used to immunize chimpanzees to determine whether the antibodies are protective. Alternatively, cells infected with the chimeric viruses of the invention may be passaged in cell culture to produce attenuated viruses which can be tested as candidate live vaccines. In assaying the ability of the chimeric viruses of the invention to infect mammals one can assay sera or liver of the infected mammal by RT-PCR to determine viral titer. In addition, the virulence phenotype of the virus produced by transfection of mammals with the sequences of the invention can be monitored by methods known in the art such as measurement of liver enzyme levels (alanine aminotransferase (ALT) or isocitrate dehydrogenase (ICD)) or by histopathology of liver biopsies.

Alternatively, mutations may be introduced into the HCV portion of the HCV/BVDV chimeras of the invention in order to enable the production of virions in cell cultures which could then be tested in vivo for improved vaccine properties.

In another embodiment, multiple chimeras containing HCV structural genes (or fragments thereof, such as the HVR1) from multiple genotypes can be administered to generate multivalent vaccines.

When used as a vaccine, the chimeric virions can be administered alone or in a suitable diluent, including, but not limited to, water, saline, or some type of buffered medium. The vaccine according to the present invention may be administered to an animal, especially a mammal, and most especially a human, by a variety of routes, including, but not limited to, intradermally, intramuscularly, subcutaneously, or in

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serum free of BVDV and antibodies to BVDV (Boyt Veterinary, Neosho, MO) was used. All cells were incubated at 37°C in 5% CO₂.

Table 1

List of Cell Lines

Cell	Origin	Medium
EBTr(A)	Embryonic bovine trachea	10% FBS/MEM
BT	Bovine turbinate	10% horse serum/MEM
MDBK	Bovine kidney	10% horse serum/MEM
EBTr (B)	Embryonic bovine trachea	10% FBS/MEM
Huh 7	human hepatoma	10% FBS/DMEM F12

Antibodies

H79: plasma from patient H obtained in the chronic phase two years after the onset of HCV infection (11); CH1530: serum pool from chimpanzee 1530, obtained in the chronic phase one to two years after the onset of HCV infection. Chimpanzee 1530 became infected with HCV following intrahepatic transfection with pCV-H77C (Yanagi 1997); LMF86 and LMF87: anti-HVR1 (Farci 1996), rabbit anti-peptide sera; Mab NS: anti-BVDV NS3 murine monoclonal antibody kindly provided by Dr. E. Dubovi (Cornell University, Ithaca, NY).

Construction of HCV/BVDV chimeric clone

The C, E1 and E2 genes originating from an infectious clone of the H77 strain of HCV (pCV-H77C, ref. Yanagi 1997), and the backbone originating from two subgenomic plasmids (pBV18-F2 and pSDMlu-3'), used by Vassilev et al. (Vassilev 1997) to generate the infectious clone of the NADL strain of BVDV (pVVNADL), were used to construct the chimeric cDNA clone pHCV-BVDV-3 (ATCC deposit Number PTA-158). The chimeric clone includes sequences corresponding to nucleotides

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HCV/BVDV sequence of the final preparation was determined using standard procedures and about 90 specific sense and antisense primers. Clone pHCV/BVDV-3 was apparently stable since the digestion pattern was as expected following retransformation. The complete sequence differed slightly from the published BVDV sequence of the NADL strain (21), but encoded an intact polyprotein.

Table 2

Oligonucleotides used for PCR amplification

Name	Sequences (5' - 3')	Underline
N-C/H77/S	CAAGTTGCAGCACGAATCCTAAACCTCAAAGAA	N OF BVDV-NADL
MluI/NADL/S	CACGCGTATCGATGAATTCTG	Mlu I
B2-P7/NADL/S	AGCGGAGGCGATTTCAGTATGGATCAGGGGAAGTG	E2 OF HCV
E2-P7/H77/R	ATACTGAATCGCCTCCGCTTGGGATATGAG	P7 OF BVDV-NADL
N-C/NADL/R	AGGATTTCGTGCTGCAACTTGTGACCCATAGAGGG	Core OF HCV
	CAGTC	
BanI/NADL/R	TACCAGGCTGAGAATGCACTGTAAC	Bsm I
2937S-HCBV	CCTTGTCCACCGGCCTCATCCACCTCCACC	
1353S-NADL	CAATTCATGGTATGATGGATGC	
1419S-NADL	AGTGGAAACAAGCATGGTTGGTG	
2335-NADL	CCACGTGGACGAGGGCATGCC	
3342R-NADL	CCTGAATCGGCCTTTACCACATCCCCAATC	
1623R-NADL	TTCTTTCTTTCTTGCAACCTGT	
1590R-NADL	GGGCTATCTCTAGCTTGTGTTAC	
389R-NADL	CCATGTGCCATGTACAGCAGAG	

Transfection of cell lines with transcribed RNA

The plasmid pHCV/BVDV-3 was linearized with SacII (NEB) and treated with T4 DNA polymerase (GIBCO/BRL) to remove the resulting 3' overhang. A truncated form of pHCV/BVDV-3, generated by digestion with HindIII, was used as a negative control. Two micrograms of DNA were transcribed at 37°C for 2 hrs in a 100 µl reaction volume containing 50 U of T7 RNA polymerase (Promega), 10 mM DTT (Promega), 120 U of Rnasin (Promega) and 1 mM rNTPs (GIBCO/BRL). Five microliters of the final reaction mixture was analyzed

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with phosphate buffered saline (PBS) for 10 min. Thereafter, cells were incubated for 20-60 min at 37°C with primary antibodies diluted in 10% bovine serum albumin (BSA) in PBS. As primary antibodies we used an anti-HCV human plasma sample (H79, 1:100 dilution), an anti-HCV chimpanzee serum (CH1530, 1:100 dilution) and an anti-BVDV NS3 monoclonal antibody (Mab-NS, 1:10 dilution). After washing with PBS for 15 min, cells were incubated for 20-40 min at 37°C with secondary antibodies; fluorescein-isothiocyanate (FITC)-conjugated goat anti-human antibody (SIGMA) for H79 and CH1530, and rhodamine-conjugated anti-mouse antibody (PIERCE) for anti-BVDV NS3. For double staining, H79 or CH1530 anti-HCV antibody was mixed with the anti-BVDV NS3 monoclonal antibody and incubated on fixed cells as above, followed by washing and incubation with a mixture of both secondary antibodies. After washing, slides were mounted and examined by fluorescence microscopy (Zeiss).

Determination of sucrose
gradient density of recovered viruses

A T150 flask of EBTr(A) cells was inoculated with virus stock. At days 9 and 13, respectively, supernatant was harvested. A total of 70 ml of supernatant was layered over 20% sucrose in TN buffer [50mM Tris and 100mM NaCl (pH 7.4)] and centrifuged at 28,000 rpm in an SW28 swinging bucket rotor (Beckman) for 19 hrs at 4°C. The pellet was resuspended in 100 µl of TN buffer. For sucrose equilibrium gradient centrifugation, the resuspended pellet was layered onto a 20-60% (wt/wt) sucrose gradient in TN buffer and centrifuged at 36,000 rpm in an SW40 swinging bucket

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incubated with ECL Western blotting detection reagent (Amersham) and exposed to film.

Detection of chimeric genomic RNA by RT-PCR assays

Total RNA was extracted with the TRIzol reagent from 10 or 100 µl of cell suspension, supernatant or material from the sucrose gradient. The RNA pellet was resuspended in 10 mM dithiothreitol (DTT) containing 5% (vol/vol) of RNAsin (20-40 U/µl) (Promega). The RT was performed with avian myeloblastosis virus reverse transcriptase (Promega) and the external anti-sense primer (see below) and PCR was performed with AmpliTaq Gold DNA polymerase (Perkin Elmer) as described (Bukh, 1998a). Specificity was confirmed by sequence analysis of selected DNA products. Each set of experiments included a low titer positive control sample and appropriate negative controls. HCV/BVDV chimeric genomes were detected in one round of PCR with the primers 2937S-HCBV and 3342R-HCBV (Table 2). The structural region of BVDV was detected in an RT-nested PCR with external primers 1353S-NADL and 1623R-NADL and internal primers 1419S-NADL and 1590R-NADL (Table 2). These primers were conserved among all known BVDV strains. Finally, the 5' UTR sequence of BVDV was detected by using universal primers that detect both HCV and BVDV (Bukh 1992, Yanagi 1996), as well as universally conserved BVDV primers (233S-NADL and 389R-NADL). The genome equivalent (GE) titer of HCV, BVDV and HCV/BVDV in positive samples was determined by RT-nested PCR on 10-fold serial dilutions of the extracted RNA (Bukh 1998a). One GE was defined as the number of genomes present in the highest dilution

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the Vectastain Elite kit (Vector Laboratories, Burlingame, CA) for peroxidase staining per the manufacturer's directions. The peroxidase substrate kit was Vector VIP (Vector Laboratory). Color development was stopped by washing the slide with water followed by air drying. Foci were counted with the aid of a dissecting microscope.

Focus neutralization assay

The assay was performed exactly as for the focus assay except the 200 μ l inoculum consisted of 100 μ l of chimeric virus diluted in 10% DMEM, 20 μ l undiluted test or control serum, and 80 μ l 10% DMEM. Each 200 μ l sample was incubated at 4° C in ice overnight prior to inoculation of cells. Sera included fetal calf serum (Boyt) and rabbit pre-immune serum as negative controls, hyperimmune rabbit antisera raised to peptides spanning the HVR1 region of the H27 strain of HCV (Farci, 1996), and goat anti-BVDV (VMRD Pullman, WA) prepared without azide. All sera had been heat-inactivated at 56° C for 30 minutes.

Immunofluoresence neutralization assay in Huh7 cells

Two hundred microliters of chimeric virus was mixed with 20 μ l of serum or plasma, incubated on ice overnight and added to one well of a four-well chamber slide. After 2 hours at 30°C, 1 ml of agarose overlay was added as for the focus assay. Four days later, slides were fixed and stained as for immunofluoresence microscopy and stained cells were manually counted by scanning the entire well using a Zeis microscope and the 40X objective.

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a particle containing significant amounts of HCV proteins.

Although the proportion of cells producing HCV proteins increased in EBTr(A) cells, it remained low in the MDBK, BT, and EBTr(B) cell lines, suggesting that the virus was not spreading in these cells. In order to determine if these cells were making infectious virus, a homologous transmission was attempted by removing supernatant from each transfected culture and adding it to a new culture of the same cell line. The only successful transmission was from the transfected EBTr(A) cells to naive EBTr(A) cells (Table 3). Therefore, although the chimeric virus genome could replicate in all four cell lines and produced HCV proteins, only in the EBTr(A) cells was virion morphogenesis coupled with availability of a receptor conducive to infection.

Table 3

Homologous passage and heterologous passage

	Transfection	Homologous passage	Heterologous passage
EBTr(A)	+	+	
EBTr(B)	+	-	+
BT	+	-	+
MDBK	+	-	+
: Supernatants from transfected cells were passed onto new cells of the same type.			
: Supernatants from transfected EBTr(A) cells were passed to indicated cells.			

Two heterologous transmission experiments were performed to determine if the three other cell lines released infectious particles. In the first experiment, supernatant from transfected MDBK cells was inoculated onto the EBTr(A) cells. Immunofluorescence microscopy

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RT-PCR primers designed to amplify known BVDV strains were able to amplify a cDNA fragment from uninoculated EBTr(A) cultures (titer: 10^6 GE/ml). The sequence of the cDNA was determined and found to match that of the CP-7 strain of BVDV (18).

Based on the data that only EBTr cells harboring BVDV were able to produce infectious particles containing the chimeric genome, it was hypothesized that the endogenous virus was serving as a helper virus, possibly by providing BVDV structural proteins. In order to determine if the infectious chimeric particles contained BVDV glycoproteins, a focus assay was developed in which cells expressing the chimeric genome were identified by their reactivity with CH1530 anti-HCV serum. An infectivity titer of 10^5 chimeric viruses/ml was obtained for passage 10 virus, which had an RT-PCR titer of 10^8 to 10^9 GE/ml. Chimeric virus produced in EBTr(A) cells was examined for its susceptibility to neutralization by anti-serum to BVDV as compared to neutralization by anti-sera raised against the hypervariable region 1 (HVR1) of the same HCV strain as was in the chimera. Dilutions of chimeric virus were incubated overnight with anti-BVDV, anti-HCV or control sera and the number of infectious particles remaining was determined by the focus assay (Table 4). The number of foci in the rabbit and bovine serum controls decreased in parallel with the dilution factor, indicating that the assay was linear and reliable. The anti-HCV sera did not neutralize the chimera. In contrast, anti-BVDV eliminated all foci at each dilution, suggesting that each and every infectious particle contained BVDV glycoproteins and that they were

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into these cells might be totally independent of HCV glycoprotein. Thirdly, the HCV E2 glycoprotein might not have folded properly to function or to be recognized by the antibody. The question of the neutralizing potential of the anti-HVR1 serum cannot be answered at this time. By an immunofluorescence microscopy assay, the anti-HVR1 serum had titers of 1:1600 and 1:3200 for rabbits LMF86 and LMF87 respectively but the antibody detected by this assay is not necessarily neutralizing antibody. The functionality of the HCV glycoproteins would best be proved by infecting cells which are not susceptible to infection by BVDV due to an absence of the BVDV receptor. Huh 7 cells were chosen as an experimental system to test for functional HCV glycoproteins because they are a human cell line which grows well and is of hepatocyte origin. Attempts to infect Huh 7 cells with the endogenous BVDV virus of the EBTr(A) cell line were not successful, suggesting either that the receptor for BVDV was absent or that the BVDV genome was unable to replicate in these cells. Attempts to infect the Huh 7 cells with the chimera were more successful. Four days after incubation with 2×10^4 EBTr(A) tissue culture infectious doses (TCID) of the chimera, Huh 7 cells could be stained with antibody to NS3 as well as with antibody to HCV. Quantification of the number of infected cells indicated that the inoculum contained 10^3 TCID /ml for Huh 7 as compared to 10^5 /ml for EBTr(A) cells. Although the cells could be infected, the virus did not spread, suggesting that in Huh 7 cells, as in the MDBK and BT cells, virions either were not assembled or were not released from cells. Most likely, the CP-7 virus could not provide the

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antibody but at a lower titer. Since the DNA vaccine expressed only the E2 glycoprotein, this protein must be involved in binding to Huh 7 cells. The plasma from chimp 1530 contained antibodies to the HCV envelope proteins as measured by ELISA or immunofluorescence microscopy but apparently, these were not neutralizing antibodies. Chimpanzee 1494 did not have demonstrable antibodies against the HCV glycoproteins so its failure to neutralize was not unexpected. Therefore, the chimera should be very useful for screening samples for neutralizing antibodies and discriminating between those that neutralize as compared to those that just bind.

Table 5

Neutralization of chimeric
virus growth in Huh 7 cells¹

Virus dilution	Number of foci ²		
	Fetal Calf Serum (Boytr)	Anti-HCV HVR1	Anti-BVDV
Undiluted	191	298	0
Dilution (1:10)	23	43	0
1. Huh 7 cells were used for infection but the virus had been grown in EBTr (A) cells.			
2. Foci stained with chimp 1530 anti-HCV and visualized by immunofluorescence microscopy.			

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glycoproteins were synthesized, it would be feasible to test purified chimeric virions as a candidate inactivated vaccine. Purified chimeric virions can be tested first in mice and if antibody to HCV is produced, the virions will be tested in chimpanzees to determine if the candidate vaccine is efficacious. The fact that virions grown in EBTr(A) cells were able to infect Huh 7 cells and were neutralized by some anti-HCV positive plasmas (Table 6) suggests that such chimeric viruses could be used to screen for neutralizing antibodies to HCV as well as to screen other cell lines for HCV receptors. The infectivity of the chimera proves the principle that HCV-BVDV chimeras can serve as a useful tool for studying the molecular biology of HCV. The glycoprotein genes from the five other genotypes of HCV can be similarly inserted into the BVDV backbone in order to provide an assay for antibodies to each genotype. Additional chimeras are being constructed in which the core protein of BVDV is included so that only the glycoproteins of HCV are introduced. If BVDV core is critical for encapsidation of the RNA, it may be possible to generate chimeric viruses in the absence of helper. It will also be revealing to determine if the HCV contribution to the chimera can be localized to either E1 or E2 alone. Such a chimera will be tested for its ability to infect EBTr(A) and Huh 7 cells. These studies will help determine the relative importance of E1 and E2 for infection of Huh 7 cells and may define any association with the BVDV glycoproteins. In addition, chimeras in which the BVDV nonstructural genes such as p7 or NS4B or NS5A are replaced with the corresponding genes of HCV may also be generated to

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-37-

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has been replaced by the non-structural region of a hepatitis C virus genome.

8. The nucleic acid molecule of claim 7, wherein at least one gene from the non-structural region of the BVDV genome has been replaced by the corresponding gene from the non-structural region of a hepatitis C virus genome.

9. A DNA construct comprising the nucleic acid molecule of claims 1, 2 or 7.

10. An RNA transcript of the DNA construct of claim 9.

11. A polypeptide encoded by the nucleic acid molecule according to claim 1.

12. A polypeptide encoded by the nucleic acid molecule according to claim 2.

13. The polypeptide according to claim 12, wherein said polypeptide is selected from the group consisting of E1, E2 or C.

14. A host cell transfected with the DNA construct of claim 9.

15. A host cell transfected with the RNA transcript of claim 10.

16. A chimeric HCV-BVDV virus produced by transfecting a host cell with the DNA construct of claim 9.

17. A chimeric HCV-BVDV virus produced by transfecting a host cell with the RNA transcript of claim 10.

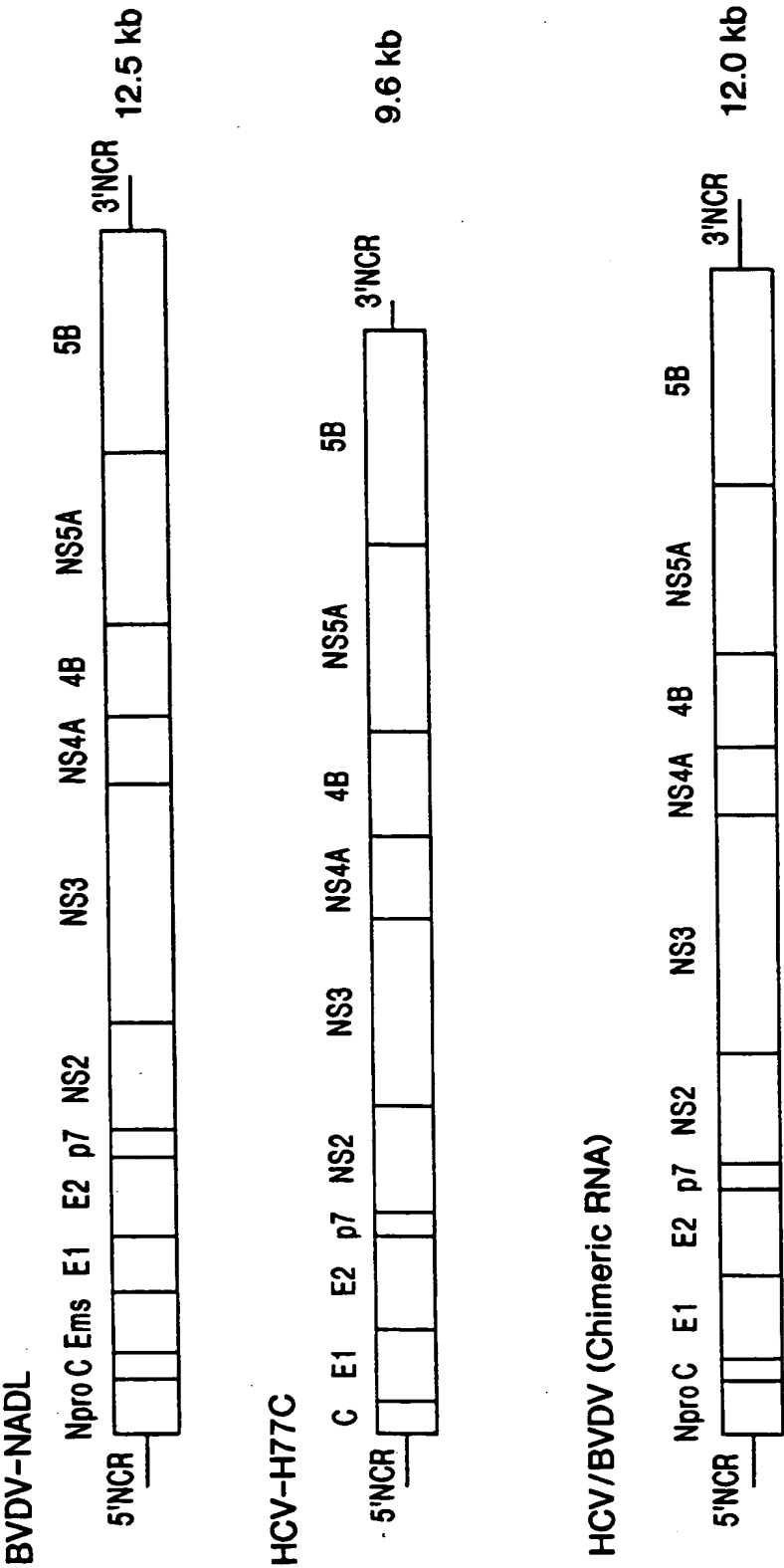


FIG. 1

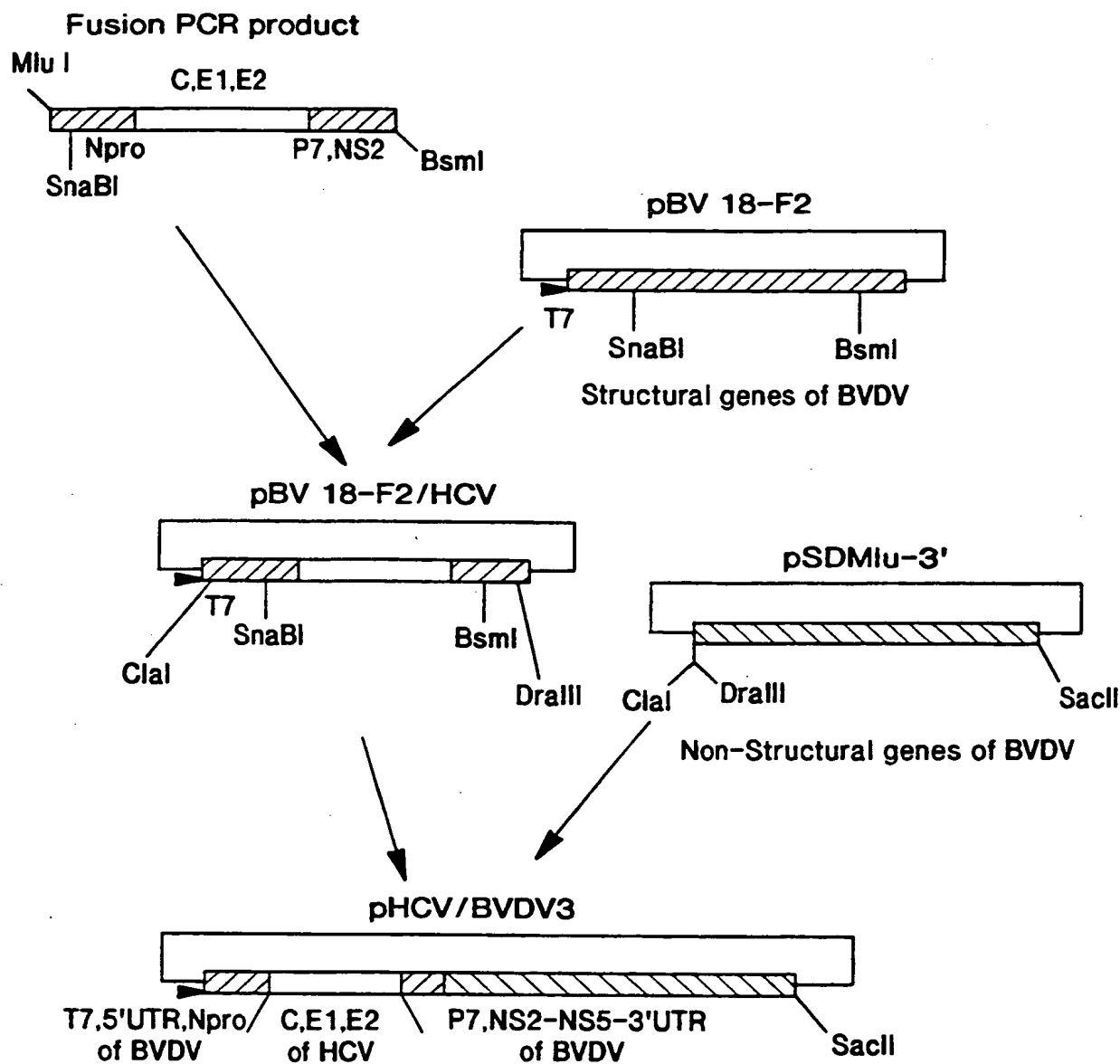


FIG. 2

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCCC	TGATGGGGGG	GACACTCCAC	CATGAATCAC	TCCCTGTGA	50
GGAACACTG	TCTTCAAGCA	GAAAGGGTCT	AGCCATGGGG	TTAGTATGAG	100
TGTCGTGCAG	CCTCCAGGAC	CCCCCCTCCC	GGGAGAGCCA	TAGTGGTCTG	150
CGGAACCGGT	GAGTACACCG	GAATTGCCAG	GACGACCGGG	TCCTTTCTTG	200
GATAAACCGG	CTCAATGCGT	GGAGATTGCG	GGGTGCCCCC	GCAAGACTGC	250
TAGCCGAGTA	GTGTGTTGGT	GCGAAAGGCC	TTGTGGTACT	GGCTGATAGG	300
GTGCTTGCGA	GTGCCCCGGG	AGGTCTGGTA	GACCGTGCAC	CATGAGCAAG	350
AATCCCTAAC	CTCAAAGAAA	AACCAAAAGT	AACACCAACC	GTCGCCCCAC	400
GGACGTCAAG	TTCCCCGGTG	GCGGTGAGAT	CGTTGGTGGG	GTTTACTTGT	450
TGCCCCGCGC	GGGCCCCAGA	TTGGGTGTGC	GCGGACGAG	GAAGACTTCC	500
GAGCGGTGCG	AACCTGAGG	TAGAAGTCAG	CCTATCCCCA	AGGCAGGTGG	550
GGCCGAGGGC	AGGACCTGGG	CTCAGCCCGG	GTACCCCTTG	CCCCCTCTAT	600
GCAATGAGGG	TTGCGGGTGG	GCGGGATGGC	TCCTGTCTCC	CCGTGGCTCT	650
CGGCCCTAGCT	GGGGCCCCAC	AGACCCCCCG	CGTAGGTGCG	GCAATTGCGG	700
TAAGGTGATC	GATACCTTCA	CGTGGCGCTT	CGCCGACCTC	ATGGGGTACA	750
TACCGCTCGT	CGGCGCCCCC	CTTGGAGGCG	CTGCCAGGGC	CCTGGCGCAT	800
GGCGTCCGGG	TTCTTGAAGA	CGCGGTGAAC	TATGCAACAG	GGAACCTTCC	850
TGGTTGCTCT	TTCTCTATCT	TCCTTCTGGC	CCTGCTCTCT	TGCTTACTGT	900
TGCCCCGCTT	AGCCTACCAA	GTGCGCAATT	CCTCGGGGCT	TTACCATGTC	950
ACCAATGATT	GGCCTAAGTC	GAGTATTGTG	TACGAGGCGG	CCGATGCCAT	1000
CCTGCACACT	CCGGGGTGTG	TCCCTTGGGT	TGCGAGGGGT	AACGCCTCGA	1050
GGTGTGTTGG	GGCGGTGACC	CCCACGGTGG	CCACCAGGGA	CGGCAAGTCT	1100
CCCACAACGC	AGCTTCGACG	TCATATCGAT	CTGCTTGTGC	GGAGCGCCAC	1150
CCCTGTGCTG	GGCCTCTACG	TGGGGGACCT	GTGCGGGTCT	GTCCTTCTTG	1200
TGGGTCAACT	GTTTACCTTC	TCTCCCAGGC	GCCACTGGAC	GACGCAAGAC	1250
TGCAATTGTT	CTATCTATCC	CGGOCATATA	ACGGGTGATC	GCATGGCATG	1300
GGATATGATG	ATGAAGTGGT	CCCCTACGGC	AGCGTTGGTG	GTAGCTCAGC	1350
TGCTCCGGAT	CCCACAAGCC	ATCATGGACA	TGATCGCTGG	TGCTCACTGG	1400
GGAGTCCCTG	CGGGCATAGC	GTATTTCTCC	ATGGTGGGGA	ACTGGGGCGA	1450
GGTCCCTGGT	GTGCTGCTGC	TATTTGCGCG	CGTGCAGCGG	GAAACCCACG	1500
TCACCGGGGG	AAATGCGGGC	CGCACCACGG	CTGGGCTTGT	TGGTCTCCTT	1550
ACACCAGGCG	CCAAGCAGAA	CATCCAAGTG	ATCAACACCA	ACGGCAGTTG	1600
GCACATCAAT	AGCACGGGCT	TGAATTGCAA	TGAAAGCCTT	AACACCGGCT	1650
GGTTAGCAGG	GCTCTTCTAT	CAACACAAAT	TCAACTCTTC	AGGCTGTGCT	1700
GAGAGGTGCG	CCAGCTGCGG	ACGCCTTACC	GATTTTGCCC	AGGGCTGGGG	1750
TCCTATCAGT	TATGCCAACG	GAAGCGGCTT	CGACGAACGC	CCCTACTGCT	1800
GGCACTACCC	TCCAAGACCT	TGTGGCATTG	TGCCCCGAAA	GAGCGTGTGT	1850
GGCCCCGTAT	ATTGCTTCAC	TCCCAGCCCC	GTGGTGGTGG	GAACGACCGA	1900

FIG. 3A

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CAGGTCGGGC	GCGCCTACCT	ACAGCTGGGG	TGCAAATGAT	ACGGATGTCT	1950
TOGTCCTTAA	CAACACCAGG	CCACCGCTGG	GCAATTGGTT	CGGTGTGACC	2000
TGGATGAACT	CAACTGGATT	CACCAAGTG	TGCGGAGGCG	CCCTGTGTGT	2050
CATCGGAGGG	GTGGGCAACA	ACAOCITGCT	CTGCCCCACT	GATTGCTTCC	2100
GCAAACATCC	GGAAGCCACA	TACTCTCGGT	GCGGCTCCGG	TCCCTGGATT	2150
ACACCCAGGT	GCATGGTUGA	CTACCCGTAT	AGGCTTTGGC	ACTATCCTTG	2200
TACCATCAAT	TACACCATAT	TCAAAGTCAG	GATGTACGTG	GGAGGGGTGG	2250
AGCACAGGCT	GGAAGCGGCG	TGCAACTGGA	CGCGGGGCGA	ACGCTGTGAT	2300
CTGGAAGACA	GGGACAGGTC	CGAGCTCAGC	CGGTGTGCTG	TGTCCACCAC	2350
ACAGTGGCAG	GTCTTTCGGT	GTCTTTTCAC	GACCCGTGCA	GCCTTGTGCA	2400
CCGGCCTCAT	CCACCTCCAC	CAGAACATTG	TGGACGTGCA	GTACTTTGAC	2450
GGGGTAGGGT	CAAGCATGCG	GTCTTGGGCC	ATTAAAGTGG	AGTACGTGCT	2500
TCTCCTGTTC	CTTCTGCTTG	CAGACGCGCG	CGTCTGCTCC	TGCTGTGTGA	2550
TGATGTTACT	CATATCCCAA	GCGGAGCGCG	CTTTGGAGAA	CCTCGTAATA	2600
CTCAATGCAG	CATCCCTGGC	CGGGACGCAC	GGTCTGTGTG	CCTTCCCTGT	2650
GTCTTCTGCG	TTTGGCGTGT	ATCTGAAGGG	TAGGTGGGTG	CCCGGAGCGG	2700
TCTACGCCCT	CTACGGGATG	TGGCCTCTCC	TCTGCTCTCT	GCTGGCGTTG	2750
CCTCAGCGGG	CATACGCACT	GGACACGGAG	GTGGCCGCGT	CGTGTGGCGG	2800
CGTGTGTCTT	GTGGGGTAA	TGGCGCTGAC	TCTGTGCGCA	TATTACAAGC	2850
GCTATATCAG	CTGGTGCATG	TGGTGGCTTC	AGTATTTTCT	GACCAGAGTA	2900
GAAGCGCAAC	TGCACGTGTG	GGTCCCCCCC	CTCAACGTCC	GGGGGGGGCG	2950
CGATGCCGTC	ATCTTACTCA	TGTGTGTAGT	ACACCCGACC	CTGGTATTTG	3000
ACATCACCAA	ACTACTCCTG	GCCATCTTGG	GACCCCTTTG	GATCTTTCAA	3050
GCCAGTTTGC	TTAAAGTCCC	CTACTTGGTG	CGCGTTCAAG	GCCMTCTCCG	3100
GATCTGCCGG	CTAGCGCGGA	AGATAGCCCG	AGGTCAATTAC	GTGCAAATGG	3150
CCATCATCAA	GTTAGGGGCG	CTTACTGGCA	CCTATGTGTA	TAACCATCTC	3200
ACCCCTCTTC	GAGACTGGGC	GCACAACGGC	CTGCGAGATC	TGGCCGTGGC	3250
TGTGGAACCA	GTGTCTTTCT	CCCGAATGGA	GACCAAGCTC	ATCACGTGGG	3300
GGGCAGATAC	CGCCGGGTGC	GGTGACATCA	TCAACGGCTT	GCCCGTCTCT	3350
GCCCGTAGGG	GCCAGGAGAT	ACTGCTTTGG	CCAGCCGACG	GAATGGTCTC	3400
CAAGGGGTGG	AGGTTGCTGG	CGCCCATCAC	GCGGTACGCC	CAGCAGACGA	3450
GAGGCCTCCT	AGGGTGTATA	ATCACAGGCC	TGACTGGCCG	GGACAAAAC	3500
CAAGTGGAGG	GTGAGGTCCA	GATCGTGTCA	ACTGCTACCC	AAACCTTCTT	3550
GGCAACGTGC	ATCAATGGGG	TATGCTGGAC	TGTCTACCCAC	GGGGCCGGAA	3600
CGAGGACCAT	CGCATCACCC	AAGGGTCTCT	TCATCCAGAT	GTATACCAAT	3650
GTGGACCAAG	ACCTTGTGGG	CTGGCCCGCT	CCTCAAGGTT	CCCGCTCATT	3700
GACACCCITG	ACCTGCGGCT	CCTCGGACCT	TACCTGGTTC	ACGAGGCACG	3750
CCGATGTCAT	TCCCGTGCCG	CGCGAGGTG	ATAGCAGGGG	TAGCCTGCTT	3800

FIG. 3B

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TGCCCCCGGC	CCATTTCCTA	CTTGAAAGGC	TCCTCGGGGG	GTCCGCTGTT	3850
GTGCCCCGCG	GGACACGCGG	TGGGOCCTATT	CAGGGCGCGG	GTGTGCACCC	3900
GTGGAGTGGC	TAAAGCGGTG	GACTTTATCC	CTGTGGAGAA	CCTAGGGACA	3950
ACCATGAGAT	CCCCGGTGT	CACGGACAAC	TCCTCTCCAC	CAGCAGTGCC	4000
CCAGAGCTTC	CAGGTGGGCC	ACCTGCATGC	TCCACCGGC	AGCGGTAAAG	4050
GCACCAAGGT	CCCGGCTGGG	TACGCAGGCC	AGGGCTACAA	GGTGTGGTG	4100
CTCAACCCCT	CTGTGTCTGC	AACGCTGGGC	TTTGGTGCTT	ACATGTCCAA	4150
GGCCCATGGG	GTTGATCCTA	ATATCAGGAC	CGGGGTGAGA	ACAATTACCA	4200
CTGGCAGGCC	CATCACGTAC	TCCACCTACG	GCAAGTTCCT	TGCGGACGGC	4250
GGGTGCTCAG	GAGGTGCTTA	TGACATAATA	ATTTGTGACG	AGTGCCACTC	4300
CACGGATGCC	ACATCCATCT	TGGGCATCGG	CATGTCTCTT	GACCAAGCAG	4350
AGACTGCGGG	GGCGAGACTG	GTGTGTCTCG	CCACTGCTAC	CCCTCCGGGC	4400
TCCGTCACTG	TGTCCCATCC	TAACATCGAG	GAGGTTCCTC	TGTCCACCAC	4450
CGGAGAGATC	CCCTTTTACG	GCAAGGCTAT	CCCCCTCGAG	GTGATCAAGG	4500
GGGGAAGACA	TCTCATCTTC	TGCCACTCAA	AGAAGAAGTG	CGACGAGCTC	4550
GCCGCGAAGC	TGGTGGCATT	GGGCATCAAT	GCCGTGGCCT	ACTACCGCGG	4600
TCTTGACGTG	TCTGTATCC	CGACCAGCGG	CGATGTGTGC	GTGTTGTGGA	4650
CCGATGCTCT	CATGACTGGC	TTTACCGCGG	ACTTCGACTC	TGTGATAGAC	4700
TGCAACACGT	GTGTCACTCA	GACAGTCGAT	TTCAGCCTTG	ACCCCTACCTT	4750
TACCATTTGAG	ACAACCACGC	TCCCCCAGGA	TGCTGTCTCC	AGGACTCAAC	4800
GCCGGGGCAG	GACTGGCAGG	GGGAAGCCAG	GCATCTATAG	ATTTGTGGCA	4850
CCGGGGGAGC	GCCCCCTCGG	CATGTTTCGAC	TGTTCCGTCC	TCTGTGAGTG	4900
CTATGACCGG	GGCTGTGCTT	GGTATGAGCT	CACGCCCCGC	GAGACTACAG	4950
TTAGGCTACG	AGCGTACATG	AACACCCCGG	GGCTTCCCGT	GTGCCAGGAC	5000
CATCTTGAAT	TTTGGGAGGG	CGTCTTTACG	GGCTCACTC	ATATAGATGC	5050
CCACTTTTIA	TCCAGACAA	AGCAGAGTGG	GGAGAACTTT	CCTTACCTGG	5100
TAGCGTACCA	AGCCACCGTG	TGCGCTAGGG	CTCAAGCCCC	TCCCCCATCG	5150
TGGGACCAGA	TGTGGAAGTG	TTTGATCCGC	CTTAAACCCA	CCCTCCATGG	5200
GCCAACACCC	CTGCTATACA	GACTGGGCGC	TGTTTAGAAT	GAAGTCACCC	5250
TGACGCACCC	AATCACCAAA	TACATCATGA	CATGCATGTC	GGCGGACCTG	5300
GAGGTGCTCA	CGAGCACCTG	GGTGCTCGTT	GGGGGGGTCC	TGGCTGCTCT	5350
GGCCGCGTAT	TGCCTGTCAA	CAGGCTGGGT	GGTCATAGTG	GGCAGGATCG	5400
TCTTGTCCGG	GAAGCCGGCA	ATTATACCTG	ACAGGGAGGT	TCTCTACCA	5450
GAGTTCGATG	AGATGGAAGA	GTGCTCTCAG	CATTACCGT	ACATCGAGCA	5500
AGGGATGATG	CTCGCTGAGC	AGTTCAAGCA	GAAGCCCTTC	GGCTCTCTGC	5550
AGACCGCGTC	CCGCCATGCA	GAGGTATATCA	CCCTGCTGT	CCAGACCAAC	5600
TGGCAGAAAC	TCGAGGTCTT	TTGGGCGAAG	CACATGTGGA	ATTTTCATCAG	5650
TGGGATACAA	TACTTGGCGG	GCCTGTCAAC	GCTGCCTGGT	AACCCCGCCA	5700

FIG. 3C
SUBSTITUTE SHEET (RULE26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TTGCTTCATT	GATGCTTTT	ACAGCTGCG	TCACCAGCCC	ACTAACCCT	5750
GGCCAAACCC	TCCTCTTCAA	CATATTGGGG	GGGTGGGTGG	CTGCCCAGCT	5800
CGCCGCCCCC	GGTGCCGCTA	CTGCCCTTGT	GGGTGCTGGC	CTAGCTGGCG	5850
CCGCCATCGG	CAGGTTGGA	CTGGGGAAGG	TCCTGCTGGA	CATTCTTGCA	5900
GGGTATGGCG	CGGGGTGGC	GGGAGCTCTT	GTAGCATTC	AGATCATGAG	5950
CGGTGAGGTC	CCCTCCAGG	AGGACCTGGT	CAATCTGCTG	CCCGCCATCC	6000
TCTGCGCTGG	AGCCCTTGTA	GTGGGTGTTG	TCTGCGCAGC	AATACTGCGC	6050
CGGCAGGTTG	GCCCGGGCGA	GGGGGCAGTG	CAATGGATGA	ACCGGCTAAT	6100
AGCCTTCGCC	TCCCGGGGGA	ACCATGTTTC	CCCCAGCCAC	TACGTGCGCG	6150
AGAGCGATGC	AGCCGCCCCG	GTCACTGCCA	TACTCAGCAG	CCTCACTGTA	6200
ACCCAGCTCC	TGAGGCGACT	GCATCAGTGG	ATAAGCTCGG	AGTGTACCC	6250
TCCATGCTCC	GGTTCCTGGC	TAGGGACAT	CTGGGACTGG	ATATGCGAGG	6300
TGCTGAGCGA	CTTTAAGACC	TGGCTGAAAG	CCAAGCTCAT	GCCACAACCTG	6350
CCTGGGATTC	CCTTTGTGTC	CTGCCAGCGC	GGGTATAGGG	GGGTCTGGCG	6400
AGGAGACGGC	ATTATGCACA	CTCGCTGCCA	CTGTGGAGCT	GAGATCACTG	6900
GACATGTCAA	AAACGGGACG	ATGAGGATCG	TGGTCTTAG	GACCTGCAGG	6950
AACATGTGGA	GTGGGACGTT	CCCCATTAAC	GCCTACACCA	CGGGCCCCCTG	6550
TACTCCCCCT	CCTGCGCCGA	ACTATAAGTT	CGCGCTGTGG	AGGGTGTCTG	6600
CAGAGGAATA	CGTGGAGATA	AGGCGGGTGG	GGGACTTCCA	CTACGTATCG	6650
GGTATGACTA	CTGACAATCT	TAAATGCCCG	TGCCAGATCC	CATCGCCCCGA	6700
ATTTTTTACA	GAATTGGACG	GGGTGCGCCT	ACACAGGTTT	GCGCCCCCTT	6750
GCAAGCCCCCT	GCTGCGGGAG	GAGGTATCAT	TCAGAGTAGG	ACTCCACGAG	6800
TACCCGGTGG	GGTGGCAATT	ACCTTGCGAG	CCCGAACCGG	ACGTAGCCGT	6850
GTGACGTCC	ATGCTCACTG	ATCCCTCCCA	TATAACAGCA	GAGGCGGCGG	6900
GGAGAAGGTT	GGCGAGAGGG	TCACCCCTTT	CTATGGCCAG	CTCCTGGCT	6950
AGCCAGCTGT	CCGCTCCATC	TCTCAAGGCA	ACTTGCACCG	CCAACCATGA	7000
CTCCCCTGAC	GCCGAGCTCA	TAGAGGCTAA	CCTCCTGTGG	AGGCAGGAGA	7050
TGGGCGGCAA	CATCACCAGG	GTGTAGTCAG	AGAACAAGT	GGTGATTCCTG	7100
GACTCCTTCG	ATCCGCTTGT	GCCAGAGGAG	GATGAGCGGG	AGGTCTCCGT	7150
ACCTGCAGAA	ATTCTGCGGA	AGTCTCGGAG	ATTGCGCCCG	GCCCTGCCCCG	7200
TCTGGGCGCG	GCCGGACTAC	AACCCCCCGC	TAGTAGAGAC	GTGGAAAAG	7250
CCTGACTACG	AACCACTGT	GGTCCATGGC	TGCCCCCTAC	CACCTCCACG	7300
GTCCCCCTCT	GTGCCCTCCG	CTCGGAAAAA	GCGTAAGGTG	GTCTCAACCG	7350
AATCAACCTT	ATCTACTGCC	TGGCCGAGC	TGCCCACCAA	AAGTTTGGC	7400
AGCTCCTCAA	CTTCCGGCAT	TACGGGCGAC	AATAAGACAA	CATCCTCTGA	7450
GCCCCCCCCCT	TCTGGCTGCC	CCCCGACTC	CGACGTGAG	TCCATATCTT	7500
CCATGCCCCC	CCTGGAGGGG	GAGCCTGGGG	ATCCGGATCT	CAGCGACGGG	7550
TCATGGTGA	CGGTACGTAG	TGGGGCCGAC	ACCGAAGATG	TGTGTGCTG	7600

FIG. 3D

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CTCAATGICT	TATTCCTGGA	CAGGCGCACT	CGTCACCCCG	TGCGCTGCGG	7650
AAGAACAAAA	ACTGCCCATC	AAGGCACTGA	GCAACTCGTT	GCTACGCCAT	7700
CACAATCTGG	TGTATTCCAC	CACTTCACGC	AGTGCTTGCC	AAAGCCAGAA	7750
GAAAGTCACA	TTTGACAGAC	TGCAAGTTCT	GGACAGCCAT	TACCAGGACG	7800
TGCTCAAGGA	GGTCAAAGCA	GCGGCGTCAA	AAGTGAAGGC	TAACTTGCTA	7850
TCCGTAGAGG	AAGCTTGCG	CCTGACGCCC	CCACATTGAG	CCAAATCCAA	7900
GTTTGGCTAT	GGGGCAAAG	ACGTCCGTTG	CCATCCGAGA	AAGGCCGTAG	7950
CCCACATCAA	CTCCGTGTGG	AAAGAOCCTC	TGGAAGACAG	TGTAACACCA	8000
ATAGACACTA	CCATCATGGC	CAAGAACGAG	GTTTTCTGCG	TTCAGCCTGA	8050
GAAGGGGGGT	CGTAAGCCAG	CTCGTCTCAT	CGTGTTCCCC	GACCTGGGCG	8100
TGCGCGTGTG	CGAGAAGATG	GCCCTGTACG	ACGTGGTTAG	CAAGCTCCCC	8150
CTGGCCGTGA	TGGGAAGCTC	CTACGGATTG	CAATACTCAC	CAGGACAGCG	8200
GGTIGAATTG	CTCGTGCAAG	CGTGGAAGTC	CAAGAAGACC	CCGATGGGGT	8250
TCTCGTATGA	TACCCGCTGT	TTTGACTCCA	CAGTCACTGA	GAGCGACATC	8300
CGTACGGAGG	AGGCAATTTA	CCAATGTTGT	GACCTGGACC	CCCAAGCCCC	8350
CGTGGCCATC	AAGTCCCTCA	CTGAGAGGCT	TTATGTTGGG	GGCCCTCTTA	8400
CCAATTCAAG	GGGGGAAAC	TGCGGCTACC	GCAGGTGCCG	CGCGAGCGGC	8450
GTAAGTACAA	CTAGCTGTGG	TAACACCCCTC	ACTTGCTACA	TCAAGGCCCG	8500
GGCAGCCTGT	CGAGCCGCAG	GGCTCCAGGA	CTGCACCATG	CTCGTGTGTG	8550
GCGACGACTT	AGTCGTTATC	TGTGAAAGTG	CGGGGGTCCA	GGAGGACGCG	8600
GCGAGCCTGA	GAGCCTTCAC	GGAGGCTATG	ACCAGGTAAT	CCGCCCCCCC	8650
CGGGGACCCC	CCACAACCAG	AATACGACTT	GGAGCTTATA	ACATCATGCT	8700
CCTCCAACGT	GTCAGTCGCC	CACGACGGCG	CTGGAAAGAG	GGTCTACTAC	8750
CTTACCCGTG	ACCCCTACAAC	CCCCCTCGCG	AGAGCCCGGT	GGGAGACAGC	8800
AAGACACACT	CCAGTCAATT	CCTGGCTAGG	CAACATAATC	ATGTTTGCCC	8850
CCACACTGTG	GGCGAGGATG	ATACTGATGA	CCCATTTCTT	TAGCGTCCCT	8900
ATAGCCAGGG	ATCAGCTTGA	ACAGGCTCTT	AACGTGTGAG	TCTACGGAGC	8950
CTGCTACTCC	ATAGAACCAC	TGGATCTACC	TCCAATCATT	CAAGACTCC	9000
ATGGCCTCAG	CGCATTTTCA	CTCCACAGTT	ACTCTCCAGG	TGAAATCAAT	9050
AGGGTGGCCG	CATGCCCTCAG	AAAACCTGGG	GTCCCGCCCT	TCCGAGCTTG	9100
GAGACACCGG	GCCCGGAGCG	TCCGCGCTAG	GCTTCTGTCC	AGAGGAGGCA	9150
GGGCTGCCAT	ATGTGGCAAG	TACCTCTTCA	ACTGGGCAGT	AAGAACAAG	9200
CTCAAACCTA	CTCCAATAGC	GGCCGCTGGC	CGGCTGGACT	TGTCCGGTTG	9250
GTTACAGGCT	GGCTACAGCG	GGGGAGACAT	TTATCACAGC	GTTCTCTCATG	9300
CCCGGCCCCG	CTGGTTCTGG	TTTTGCCCTAC	TCCGTGCTGC	TGCAGGGGTA	9350
GGCATCTACC	TCTTCCCCAA	CCGATGAAGG	TTGGGGTAAA	CACTCCGGCC	9400
TCTTAAGCCA	TTTCCGTGTT	TTTTTTTTTT	TTTTTTTTTT	TTTTTCTTTT	9450
TTTTTTTCTT	TCCTTTCCTT	CTTTTTTTTC	TTTCTTTTTC	CCTTCTTAA	9500

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TGGTGGCTCC	ATCTTAGCCC	TAGTCACGGC	TAGCTGTGAA	AGGTCCGIGA	9550
GCCGCATGAC	TGCAGAGAGT	GCTGATACTG	GCCTCTCTGC	AGATCATGT	9599

FIG. 3F

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
MSINPKPQRK	TKRNINRRPQ	DVKFPGGGQI	VGGVYLLPFR	GPRLGVRATR	50
KTSESRQPRG	RRQPIPKARR	PEGRTWAQPG	YFWPLYGNEG	CGWAGWLLSP	100
RGSRPSWGPT	DFRRRSRNLG	KVIDTILTOGF	ADLMGYIPLV	GAPLGGAARA	150
LAHGVRVLED	GVNYATGNLP	GCSFSIFLLA	LLSCLTVPAS	AYQVRNSSL	200
YHVINDCHNS	SIVYEAADAI	LHTPGCVPCV	REGNASROW	AVTPTVATRD	250
GKLPITQLRR	HIDLLVGSAT	LCSALYVGD	CGSVFLVGQL	FTFSRRRHW	300
TQDNCSTYP	GHTTGHMAW	IMMNWSPTA	ALVVAQLLRI	PQAIMDMIAG	350
AHWGVLGIA	YFSMVGNWAK	VLVLLLFAG	VDAETHVTGG	NAGRTTAGLV	400
GLLTPGAKQN	IQLININGSW	HINSTALNCH	ESLNTGWLAG	LFYQHKFNSS	450
GCPERLASCR	RLTDFAGQWG	PISYANGSGL	DERPYCWHYP	PRPGITVPAK	500
SVCGPVYCF	PSPVVVGTTD	RSGAPTYSWG	ANDIDVFLN	NTRPPLGNWF	550
GCTWMNSTGF	TKVCGAPPCV	IGGVGNITLL	CPTDCFRKHP	EATYSRCGSG	600
PWITPRQWD	YPYRLWHYPC	TINYTIFKVR	MYVGGVEHRL	EACNWTIRGE	650
RCDLEDRDRS	ELSPLLLSTT	QWQVLPSCFT	TLPALSTGLI	HLHQNTVDVQ	700
YLYGVGSSIA	SWAIKWEYV	LLFLLLLADAR	VCSCILWMLL	ISQAEAALEN	750
LVIILNAASLA	GIHGLVSFLV	FFCFAWYLKG	RWVPGAVYAL	YGMWPLLLLL	800
LALPQRAYAL	DTEVAASCGG	VVLVGLMALT	LSPYYKRYIS	WOMWMLQYFL	850
TRVEAQLHW	VPPLNVRGGR	DAVILLMCVV	HPITLVFDITK	LLLAIFGPLW	900
ILQASLLKVP	YFVRVQGLLR	ICALARKIAG	GHYVQMAIHK	LGALTGTIVY	950
NHLTPLRLWA	HNGLRDLAVA	VEPVVFSRME	TKLITWGADT	AACGDIINGL	1000
PVSARRQGEI	LLGPADGMVS	KGRLLAPIT	AYAQQTRGLL	GCIITSLTGR	1050
DKNQVEGEVQ	IVSTATQTFL	ATCINGVCWT	VYHAGIRTI	ASPKGPFVIQ	1100
YTNVDQDLVG	WPAPOGSRSL	TPCTCGSSDL	YLVIRHADVI	PVRRRGDSRG	1150
SLLSPRPISY	LKGSSGGPLL	CPAGHAVGLF	RAAVCTRGVA	KAVDFIPVEN	1200
LGTIMRSPVF	TDNSSPPAVP	QSFQVAHLHA	PTGSGKSTKV	PAAYAAQGYK	1250
VLVLNPSVAA	TLGFGAYMSK	AHGVDENIRT	GVRTTTTGSP	ITYSTYGFKL	1300
ADGGCSGGAY	DIIICDECHS	TDATSIILGIG	TVLDQAETAG	ARLWLATAT	1350
PPGSVIVSHP	NIEEVALSTT	GEIPFYGKAI	PLEVIKGRH	LIFCHSKKCK	1400
DELAALKVAL	GINAVAYYRG	LDSVIPTSG	DVVVSTIDAL	MIGFTGDFDS	1450
VIDCNCVTQ	TVDFSLOPTF	TIETTTLPQD	AVSRTQRRGR	TGRGKPGIYR	1500
FVAPGERPSG	MFDSSVLCEC	YDAGCAWYEL	TPAETTVRLR	AYMNTFGLPV	1550
CQDHLEFWEG	VFTGLTHIDA	HFLSQTKQSG	ENFPYLVAYQ	ATVCARAQAP	1600
PPSWDQMKC	LIRLKPTLHG	PTFLLYRLGA	VQNEVTLTHP	ITKYIMTQMS	1650
ADLEVVTSTW	VLVGGVLAAL	AAYCLSTGCV	VIVGRIVLSG	KPAIIPDREV	1700
LYQEFDEMEE	CSQHLPIYEQ	GMLAEQFKQ	KALGLLQIAS	RHAEVITPAV	1750
QTNWQKLEVF	WAKHMANFIS	GIQYLAGLST	LPGNPAIASL	MAFTAAVTSP	1800
LTTGQTLLEN	ILGGWAAQL	AAPGAATAFV	GAGLAGAAIG	SVGLGKVLVD	1850
ILAGYGAGVA	GALVAFKIMS	GEVPSTEDLV	NLLPAILSPG	ALVVGWVCAA	1900

FIG. 3G
SUBSTITUTE SHEET (RULE26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ILRRHVGPGE	GAVQWMNRLI	AFASRGNHVS	PTHVVPESDA	AARVTAILSS	1950
LTVTQLLRRL	HQWISSECTT	PCSGSWLRDI	WDWICEVLSD	FKTWLKAKLM	2000
PQLFGIPFVS	CQRGYRGWR	GCGIMHIRCH	CGAETTGHVK	NGIMRIVGPR	2050
TCRNMWSGTF	PINAYTTGPC	TPLPARNYKF	ALWRVSAEEY	VEIRRVGDFH	2100
YVSGMTIDNL	KCPQQIPSPE	FFTELDGVRL	HRFAPCKPL	LREEVSFRVG	2150
LHEYFVGSQI	PCEPEPDVAV	LTSMLTDPSH	ITAEAGRRL	ARGSPPSMAS	2200
SSASQLSAPS	LKATCTANHD	SPDAELIEAN	LLWRQEMGGN	ITRVESENKV	2250
VILDSFDPLV	AEEDEREVS	PAEILRKSRR	FARALPWAR	PDYNPFLVET	2300
WKKPDYEPPV	VHGCPLPPPR	SPFVPPPRKK	RTVVLTESTL	STALAEIATK	2350
SFGSSSTSGI	TGINTTTSSE	PAPSGCPPDS	DVESYSSMPP	LEGEFGDPDL	2400
SDGSWSIVSS	GADTEDVCC	SMSYSWIGAL	VTPCAAEEQK	LPINALSNSL	2450
LRHNLVYST	TSRSACQROK	KVTFDRLQVL	DSHYQDLKE	VKAAASKVKA	2500
NLLSVEEACS	LTPPHSAKSK	FGYGAKDVR	HARKAVAHIN	SWKDLLED	2550
VTPIDTTIMA	KNEVFCVQPE	KGGRKPARLI	VFPDLGVRVC	EKMALYDVS	2600
KLPLAVMGSS	YGFQYSPGQR	VEFLVQAWKS	KKTPMGFSYD	TRCFDSTVTE	2650
SDIRTEEATY	QCCDLDPQAR	VAIKSLTERL	YVGGLINSR	GENCGYRRCR	2700
ASGVLTTSCG	NILTCYIKAR	AACRAAGLQD	CTMLVCGDDL	VVICESAGVQ	2750
EDAASLRAFT	EAMTRYSAAP	GDPPQPEYDL	ELITSCSSNV	SVAHDGACKR	2800
VYYLTRDPTT	FLARAAWETA	RHTPVNSWLG	NIIMFAPTLW	ARMILMIHFF	2850
SVLIARDQLE	QALNCETYGA	CYSIEPLDLP	PIIQRLHGLS	AFSLHSYSEG	2900
EINRVAACLR	KLGVPPLRAW	RHRARSVRAR	LLSRGGRAAI	CGKYLEFNWAV	2950
RTKLKLTPIA	AAGRDLDSGW	FTAGYSGGDI	YHSVSHARPR	WFWFCLLLLA	3000
AGVGITYLLPN	R				3011

FIG. 3H

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCCC	TGATGGGGGC	GACACTCCAC	CATGAATCAC	TCCCCGTGTA	50
GGAACACTG	TCTTCACGCA	GAAAGCGTCT	AGCCATGGCG	TTAGTATGAG	100
TGTCTGTGAG	CCTCCAGGAC	CCCCCCTCCC	GGGAGAGCCA	TAGTGGTCTG	150
CGGAACCGGT	GAGTACACCG	GAATTGCCAG	GACGACCGGG	TCCCTTCTTG	200
GATCAACCCG	CTCAATGCCT	GGAGATTGCG	GCGTGCCCCC	GCGAGACTGC	250
TAGCCGAGTA	GTGTTGGGTC	GCGAAAGGCC	TTGTGGTACT	GCCGTGATAGG	300
GTGCTTGCGA	GTGCCCCGGG	AGGTCTCGTA	GACCGTGCAC	CATGAGCAGG	350
AATCCTAAAC	CTCAAAGAAA	AACCAAACGT	AACACCAACC	GCCGCCACAA	400
GGACGTCAAG	TTCCCCGGCG	GTGGTCAGAT	CGTTGGTGGG	GTTTACCTGT	450
TGCCGCGCAG	GGGCCCCAGG	TTGGGTGTGC	GCGCGACTAG	GAAGGCTTCC	500
GAGCGGTGCG	AACCTCGTGG	AAGGCGACAA	CCTATCCCAA	AGGCTCGCCG	550
ACCCGAGGGC	AGGGCCTGGG	CTCAGCCCCG	GTACCCCTTG	CCCCCTATG	600
GCAATGAGGG	CCTGGGGTGG	GCAGGATGGC	TCTGTGACCC	CCGCGGCTCC	650
CGGCCTAGTT	GGGGCCCCAC	GGACCCCCCG	CGTAGGTGCG	GTAACCTGGG	700
TAAGGTCATC	GATACCCCTA	CATGCGGCTT	CGCCGATCTC	ATGGGGTACA	750
TTCCGCTCGT	CGGCGCCCCC	CTAGGGGGCG	CTGCCAGGGC	CTTGGCACAC	800
GGTGTCCGGG	TTCTGGAGGA	CGGCGTGAAC	TATGCAACAG	GGAACTTGCC	850
CGGTGTCTCT	TTCTCTATCT	TCTCTTTGGC	TCTGTGTGCC	TGTTTGACCA	900
TCCCAGCTTC	CGCTTATGAA	GTGCGCAACG	TGTCCGGGAT	ATAACCATGTC	950
ACGAACGACT	GCTCCAACTC	AAGCATTGTG	TATGAGGCAG	CGGACGTGAT	1000
CATGCATACT	CCCGGGTGCG	TGCCCTGTGT	TCAGGAGGGT	AACAGCTCCC	1050
GTGTCTGGGT	AGCGCTCACT	CCCACGCTCG	CGGCCAGGAA	TGCCAGCGTC	1100
CCCACTACGA	CAATACGACG	CCACGTGAC	TTGCTCGTTG	GGACGGCTGC	1150
TTTCTGTCTC	GCTATGTACG	TGGGGGATCT	CTGCCGATCT	ATTTTCCCTC	1200
TCTCCCAGCT	GTTCACCTTC	TGCTCTCGCC	GGCATGAGAC	AGTGCAGGAC	1250
TGCAACTGCT	CAATCTATCC	CGGCCATGTA	TCAGGTACCC	GCATGGCTTG	1300
GGATATGATG	ATGAACGTGT	CACCTACAAC	AGCCCTAGTG	GTGTCCGAGT	1350
TGCTCCGGAT	CCCACAAGCT	GTGCTGGACA	TGGTGGCGGG	GGCCCACTGG	1400
GGAGTCCCTG	CGGCGCTTGC	CTACTATTCC	ATGGTAGGGA	ACTGGGCTAA	1450
GGTTCTGATT	GTGCGCTTAC	TCTTTGCGCG	CGTTGACGGG	GAGACCCACA	1500
CGACGGGGAG	GGTGGCGGGC	CACACCACCT	CCGGGTTCAC	GTCCCTTTTC	1550
TCATCTGGGG	CGTCTCAGAA	AATCCAGCTT	GTGAATACCA	ACGGCAGCTG	1600
GCACATCAAC	AGGACTGCCC	TAAATTGCAA	TGACTCCCTC	CAAACCTGGT	1650
TCTTTGCGGC	GCTGTTTTAC	GCACACAAGT	TCAACTCGTC	CGGGTGCCCC	1700
GAGCGCATGG	CCAGCTGCGG	CCCCATTGAC	TGGTTGCCCC	AGGGGTGGGG	1750
CCCCATCACC	TATACTAAGC	CTAACAGCTC	GGATCAGAGG	CCTTATTGCT	1800
GGCATTACGC	GCCTCGACCG	TGTGGTGTGC	TACCCGCGTC	GCAGGTGTGT	1850
GGTCCAGTGT	ATTGTTTCAC	CCCAAGCCCT	GTGTGTGGTG	GGACCACCGA	1900

FIG. 4A

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCGTTCCGGT	GTCCCTACGT	ATAGCTGGGG	GGAGAATGAG	ACAGACGTGA	1950
TGCTCCTCAA	CAACACGGGT	CCGCCACAAG	GCAACTGGTT	CGGCTGTACA	2000
TGGATGAATA	GTACTGGGTT	CACTAAGACG	TGCGGAGGTC	CCCCGIGTAA	2050
CATCGGGGGG	GTCGGTAACC	GCACCTTGAT	CTGCCCCAAG	GA CTGCTTCC	2100
GGAAGCACCC	CGAGGCTACT	TACACAAAAT	GTGGCTCGGG	GCCCTGGTTG	2150
ACACCTAGGT	GCCTAGTAGA	CTACCCATAC	AGGCTTTGGC	ACTACCCCTG	2200
CACTCTCAAT	TTTTCCATCT	TTAAGGTTAG	GATGTATGTG	GGGGCGGTGG	2250
AGCACAGGCT	CAATGCCGCA	TGCAATTGGA	CTCGAGGAGA	GCGCTGTAAAC	2300
TTGGAGGACA	GGGATAGGTC	AGAACTCAGC	CCGCTGCTGC	TGTCCTACAAC	2350
AGAGTGGCAG	ATACTGCCCC	GTGCTTTTAC	CACCCCTACCG	GCTTTATCCA	2400
CTGGTTTGAT	CCATCTCCAT	CAGAACATCG	TGGACGTGCA	ATACCTGTAC	2450
GGTGTAGGGT	CAGCGTTTGT	CTCCTTTTGA	ATCAAATGGG	AGTACATCCT	2500
GTTGCTTTTC	CTTCTCCTGG	CAGACGCGCG	CGTGTGTGOC	TGCTTGTGGA	2550
TGATGCTGCT	GATAGCCGAG	GCTGAGGCGG	CCTTAGAGAA	CTTGGTGGTC	2600
CTCAATGCCG	CGTCCGTGGC	CGGAGCGCAT	GGTATTCTCT	CCTTTCTTGT	2650
GTTCTTCTGC	GCCGCCCTGT	ACATTAAAGG	CAGGCTGGCT	CCTGGGGCGG	2700
CGTATGCTTT	TTATGGCGTA	TGGCCGCTGC	TCCTGCTCCT	ACTGGCGTFA	2750
CCACCACGAG	CTTACGCCCT	GGACCGGGAG	ATGGCTGCAT	CGTGCGGGGG	2800
TGCGGTCTCT	GTAGGTCTGG	TATTCTTGAC	CTTGTACCCA	TACTACAAAG	2850
TGTTTCTCAC	TAGGCTCATA	TGGTGGTTAC	AATACTTTAT	CACCAGAGCC	2900
GAGGCGCACA	TGCAAGTGTG	GGTCCCCCCC	CTCAACGTTT	GGGGAGGCGG	2950
CGATGCCATC	ATCCTCCTCA	CGTGTGCGGT	TCATCCAGAG	TTAATTTTGT	3000
ACATCACCAA	ACTCCTGCTC	GCCATACTCG	GCCCGCTCAT	GGTGCTCCAG	3050
GCTGGCATAA	CGAGAGTGCC	GTACTTGGTG	CGCGCTCAAG	GGCTCATTCG	3100
TGCATGCATG	TTAGTGCGAA	AAGTGGCCGG	GGGTCAATTAT	GTCCAAATGG	3150
TCTTCATGAA	GCTGGGCGCG	CTGACAGGTA	CGTACGTTTA	TAACCATCTT	3200
ACCCCACTGC	GGGACTGGGC	CCACGCGGGC	CTACGAGACC	TTGCGGTGGC	3250
GGTAGAGCCC	GTCGTCTTCT	CCGCCATGGA	GACCAAGGTC	ATCACCTGGG	3300
GAGCAGACAC	CGCTGCGTGT	GGGGACATCA	TCTTGGGTCT	ACCCGTCTCC	3350
GCCCGAAGGG	GGAAGGAGAT	ATTTTTTGGG	CCGGCTGATA	GTCTCGAAGG	3400
GCAAGGGTGG	CGACTCCTTG	CGCCCATCAC	GGCCTACTCC	CAACAAACGC	3450
GGGGCGTACT	TGGTTGCATC	ATCACTAGCC	TCACAGGCGG	GGACAAGAAC	3500
CAGGTGGAAG	GGGAGGTTCA	AGTGGTTTCT	ACCGCAACAC	AATCTTTTCT	3550
GGGACCTGTC	ATCAACGGCG	TGTGCTGGAC	TGTCCTACCAT	GGCGCTGGCT	3600
CGAAGACCCCT	AGCCGGTCCA	AAAGGTCCAA	TCACCCAAAT	GTACACCAAT	3650
GTAGACCTGG	ACCTCGTCCG	CTGGCAGGCG	CCCCCGGGGG	CGCGCTCCAT	3700
GACACCATGC	AGCTGTGGCA	GCTCGGACCT	TTACTTGGTC	ACGAGACATG	3750
CTGATGTCAT	TCCGGTGCCG	CGGCGAGGCG	ACAGCAGGGG	AAGTCTACTC	3800

FIG. 4B
SUBSTITUTE SHEET (RULE26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCCCCCAGGC	CCGTCTCCTA	CCTGAAAGGC	TCCTUGGGTG	GTCCATTGCT	3850
TTGCCCCCTCG	GGGCACGTCC	TGGGGCGTCT	CCGGGCTGCT	GTGTCCACCC	3900
GGGGGGTCCG	GAAGGCGGTG	GACTTTCATC	CCGTTGAGTC	TATGGAAACT	3950
ACCATGCGGT	CTCCGGTCTT	CACAGACAAC	TCAACCCCCC	CGGCTGTACC	4000
GCAGACATTC	CAAGTGGCAC	ATCTGCACGC	TCCTACTGGC	AGCGGCAAGA	4050
GCACCAAGT	GCCGGCTGGG	TATGCAGCCC	AAGGGTACAA	GGTGCTGGTC	4100
CTGAACCCGT	CCGTTGCGGC	CACCTTAGGG	TTTGGGGGGT	ATATGTCCAA	4150
GGCACACGGT	ATCGACCCTA	ACATCAGAAC	TGGGGTAAAG	ACCATTAACA	4200
CGGGCGGCTC	CATTACGTAC	TCCACCTATG	GCAAGTTCTT	TGCCGACGGT	4250
GGCTGTCTCG	GGGGCGCCTA	TGACATCATA	ATATGTGATG	AGTGCCACTC	4300
AACTGACTCG	ACTACCATCT	TGGGCATCGG	CACAGTCTTG	GACCAAGCGG	4350
AGACGGCTGG	AGCGCGGCTC	GTGCTGCTCG	CCACCGCTAC	ACCTCCGGGA	4400
TCGGTTACCG	TGCCACACCC	CAATATCGAG	GAAATAGGCC	TGTCCAACAA	4450
TGGAGAGATC	CCCTTCTATG	GCAAAGOCAT	CCCATTTGAG	GCCATCAAGG	4500
GGGGGAGGCA	TCTCATTTTC	TGCCATTCCA	AGAAGAAATG	TGACGAGCTC	4550
GCCGCAAGC	TGACAGGCCT	CGGACTGAAC	GCTGTAGCAT	ATTACCGGGG	4600
CCTTGATGTG	TCCGTTCATC	CGCCTATCGG	AGACGTCGTT	GTGCTGGCAA	4650
CAGACGCTCT	AATGACGGGT	TTCACCGGCG	ATTTTGACTC	AGTGATCGAC	4700
TGCAATACAT	GTGTACCCCA	GACAGTCGAC	TTCAGCTTGG	ATCCACCTTT	4750
CACCATTTGAG	ACGACGACCG	TGCCCCAAGA	CGCGGTGTCC	CGCTCGCAAC	4800
GGCGAGGTAG	AACTGGCAGG	GGTAGGAGTG	GCATCTACAG	GTTTGTGACT	4850
CCAGGAGAAC	GGCCCTCGGG	CATGTTTCGAT	TCTTCGGTCC	TGTGTGAGTG	4900
CTATGACCGG	GGCTGTGCTT	GGTATGAGCT	CACGCCCGCT	GAGACCTCGG	4950
TTAGGTTGCG	GGCTTACCTA	AATACACCAG	GGTTGCCCGT	CTGCCAGGAC	5000
CATCTGGAGT	TCTGGGAGAG	CGTCTTCACA	GGCCTCACCC	ACATAGATGC	5050
CCACTTCCTG	TCCCAGACTA	AACAGGCAGG	AGACAACCTT	CCTTACCCTGG	5100
TGGCATATCA	AGCTACAGTG	TGCGCCAGGG	CTCAAGCTCC	ACCTCCATCG	5150
TGGGACCAAA	TGTGGAAGTG	TCTCATACGG	CTGAAACCTA	CACTGCACGG	5200
GCCAACACCC	CTGCTGTATA	GGCTAGGAGC	CGTCCAAAAT	GAGGTTCATC	5250
TCACACACCC	CATAACTAAA	TACATCATGG	CATGCATGTC	GGCTGACCTG	5300
GAGGTCTGTA	CTAGCACCTG	GGTGCTGGTA	GGCGGAGTCC	TTGCAGCTTT	5350
GGCCGCATAC	TGCCTGACGA	CAGGCAGTGT	GGTCATTGTG	GGCAGGATCA	5400
TCTTGTCGGG	GAAGCCAGCT	GTGCTTCCCG	ACAGGGAAGT	CCTCTACCAG	5450
GAGTTCGATG	AGATGGAAGA	GTGTGCCCTA	CAACTTCCTT	ACATCGAGCA	5500
GGGAATGCAG	CTCGCCGAGC	AATTCAAGCA	AAAGGCGCTC	GGGTGTGTGC	5550
AAACGGCCAC	CAAGCAAGCG	GAGGCTGCTG	CTCCCGTGGT	GGAGTCCAAG	5600
TGGCGAGCCC	TTGAGACCTT	CTGGGCGAAG	CACATGTGGA	ATTTCATCAG	5650
CGGAATACAG	TACCTAGCAG	GCTTATCCAC	TCTGCCCTGA	AACCCCGCGA	5700

FIG. 4C

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TAGCATCATT	GATGGCATT	ACAGCTTCTA	TCTAGTCCC	GCTCACCACC	5750
CAAAACACCC	TCTGTCTTAA	CATCTTGGGG	GGATGGGTGG	CTGCCCCACT	5800
CGCTCCTCCC	AGCGCTGCGT	CAGCTTTCTG	GGCGCGCGGC	ATGCGCGGAG	5850
CGGCTGTGTG	CAGCATAGGC	CTTGGGAAGG	TGCTCGTGGG	CATCTTGGCG	5900
GGCTATGGGG	CAGGGGTAGC	CGGCGCACTC	GTGGCCTTTA	AGGTCAATGAG	5950
CGGCGAGGTG	CCCTCCACCG	AGGAOCTGGT	CAACTTACTC	OCTGCCATCC	6000
TCTCTCCTGG	TGCCCCGTGC	GTGGGGGTGG	TGTGGGCAGC	AATACTGGGT	6050
CGGCACGTGG	GCCCCGGAGA	GGGGCGTGTG	CAGTGGATGA	ACCGGCTGAT	6100
AGCGTTUGCT	TGCGGGGGTA	ACCAOCTCTC	CCCTAOCAC	TATGTGCTTG	6150
AGAGCGACGC	TGCAGCACGT	GTCACTCAGA	TCTCTCTTAG	CCTTAACATC	6200
ACTCAACTGC	TGAAGGGGCT	CCACCGATGG	ATTATATGAG	ACTGCTCTAC	6250
GCCATGCTCC	GGCTCGTGGC	TAAGGGATGT	TTGGGATTGG	ATATGCACGG	6300
TGTTGACTGA	CTTCAAGACC	TGGCTCCAGT	CCAAACTCCT	GCGCGGGTTA	6350
CCGGGAGTCC	CTTCTCTGTC	ATGCCAACGC	GGGTACAAGG	GAGTCTGGCG	6400
GGGGGACGGC	ATCATGCAAA	CCACCTGCCC	ATGCGGAGCA	CAGATCGCCG	6450
GACATGTCAA	AAACGGTTCC	ATGAGGATCG	TAGGGCCTAG	AACCTGCAGC	6500
AACACGTGGC	ACGGAAOCTT	CCCCATCAAC	GCATACACCA	CGGGACCTTG	6550
CACACCTTCC	CCGGCGCCCC	ACTATTCCAG	GGCGCTATGG	CGGGTGGCTG	6600
CTGAGGAGTA	CGTGGAGGTT	ACGCGTGTGG	GGGATTTCOA	CTACGTGACG	6650
GGCATGACCA	CTGACAACGT	AAAGTGCCCC	TGCCAGGTTT	CGGCCCCCGA	6700
ATTCTTTCAG	GAGGTGGATG	GAGTGCGGTT	GCACAGGTAC	GCTCCGGCGT	6750
GCAAACCTCT	TCTACGGGAG	GACGTACAGT	TCCAGGTCCG	GCTCAACCAA	6800
TACTTGGTGG	GGTCCGAGCT	CCCATGCGAG	CCCGAACCGG	ACGTAAACAGT	6850
GCTTACTTCC	ATGCTCACCG	ATCCCTCCCA	CATTACAGCA	GAGACGGCTA	6900
AGCGTAGGCT	GGCTAGAGGG	TCTCCCCCCT	CTTTAGCCAG	CTCATCAGCT	6950
AGCCAGTTGT	CTGCGCCTTC	TTTGAAGGCG	ACATGCACTA	CCCAACATGA	7000
CTCCCCGGAC	GCTGACCTCA	TGAGGGCCAA	CCTCTTGTGG	CGGCAGGAGA	7050
TGGGCGGAAA	CATCACTCGC	GTGGAGTCAG	AGAATAAGGT	AGTAATTCTG	7100
GACTCTTTTG	AACCGCTTCA	CGCGGAGGGG	GATGAGAGGG	AGATATCCGT	7150
CGCGGCGGAG	ATCCTGCGAA	AATCCAGGAA	GTTCCTCTCA	GCGTTGCCCA	7200
TATGGGCACG	CCCGGACTAC	AATCCTCCAC	TGCTAGAGTC	CTGGAAGGAC	7250
CCGGACTACG	TCCCTCCGGT	GGTACACCGA	TGCCCATTGC	CACTACCAA	7300
GGCTCCTCCA	ATACCACCTC	CACGGAGAAA	GAGGACGGTT	GTCTGACAG	7350
AATCCAATGT	GTCTTCTGCC	TTGGCGGAGC	TGCGCACTAA	GACCTTCGGT	7400
AGCTCCGGAT	CGTCCGCCGT	TGATAGCGGC	ACGGCGACCG	CCCTTCTCTA	7450
CCTGGCCTCC	GACGACGGTG	ACAAAGGATC	CGACGTTCAG	TGCTACTCCT	7500
CCATGCCCCC	CCTTGAAGGG	GAGCCGGGGG	ACCCCGATCT	CAGCGACGGG	7550
TCTTGGTCTA	CCGTGAGTGA	GGAGGCTAGT	GAGGATGTGG	TCTGCTGCTC	7600

FIG. 4D
SUBSTITUTE SHEET (RULE26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
AATGTCTTAT	ACGTGGACAG	GCGCCCTGAT	CACGCCATGC	GCTGCGGAGG	7650
AAAGTAAGCT	GCCCATCAAC	COGTTGAGCA	ACTCTTTGCT	GCGTCACAC	7700
AACATGGTCT	ACGCCACAAC	ATCCCGCAGC	GCAAGCCTCC	GCCAGAAGAA	7750
GGTCACCTTT	GACAGATTGC	AAGTCTGGA	TGATCATTAC	CGGGAAGTAC	7800
TCAAGGAGAT	GAAGGCGAAG	GCGTCCACAG	TTAAGGCTAA	GCTTCTATCT	7850
ATAGAGGAGG	CCTGCAAGCT	GACGCCCCCA	CATTGGGACA	AATCCAAATT	7900
TGGCTATGGG	GCAAAGGACG	TCGGGAACCT	ATCCAGCAGG	GCCTTTAACC	7950
ACATCCGCTC	CGTGTGGGAG	GACTTGCTGG	AAGACACTGA	AACACCAATT	8000
GACACCACCA	TCATGGCAAA	AAGTGAGGTT	TTCTGGGTCC	AACCAGAGAA	8050
GGGAGGCCGC	AAGCCAGCTC	GCCTTATCGT	ATTCCAGAC	CTGGGAGTTC	8100
GTGTATGCCA	GAAGATGGCC	CTTTACGACG	TGGTCTCCAC	CCTTCTCTAG	8150
GCCGTGATGG	GCTCTCATA	CGGATTTCAA	TACTCCCCCA	AGCAGCGGGT	8200
CGAGTTCTTG	GTGAATACCT	GGAATCAAA	GAAATGCOCT	ATGGGCTTCT	8250
CATATGACAC	CCGCTGTTTT	GACTCAACGG	TCACTGAGAG	TGACATTCTG	8300
GTTGAGGAGT	CAATTACCA	ATGTTGTGAC	TTGGCCCCCG	AGGCCAGACA	8350
GGCCATAAGG	TGCTCACAG	AGCGGCTTTA	CATCGGGGGT	CCCCTGACTA	8400
ACTCAAAAGG	GCAGAACTGC	GGTTATCGCC	GGTGCCGGGC	AAGTGGCGTG	8450
CTGACGACTA	GCTGCGGTAA	TACCTTCACA	TGTTACTTTGA	AGGCCACTGC	8500
AGCCTGTCCA	GCTGCAAAGC	TCCAGGACTG	CACGATGCTC	GTGAACGGAG	8550
ACGACCTTGT	CGTTATCTGT	GAAAGCGCGG	GAACCCAGGA	GGATGCGGGC	8600
GCCCTACGAG	CCTTCACGGA	GCCTATGACT	AGGTATTCCG	CCCCCCCCCG	8650
GGATCCGCCC	CAACCAGAAT	ACGACCTGGA	GCTGATAACA	TCATGTTCTT	8700
CCAATGTGTC	AGTCGCGCAC	GATGCATCTG	GCAAAAGGGT	ATACTACCTC	8750
ACCCGTGACC	CCACCACCCC	CCTTGCACGG	GCTGCGTGGG	AGACAGCTAG	8800
ACACACTCCA	ATCAACTCTT	GGCTAGGCAA	TATCATCATG	TATGCGCCCA	8850
CCCTATGGGC	AAGGATGATT	CTGATGACTC	ACTTTTCTTC	CATCCTTCTA	8900
GCTCAAGAGC	AACTTGAAAA	AGCCCTGGAT	TGTCAGATCT	ACGGGGCTTG	8950
CTACTCCATT	GAGCCACTTG	ACCTACCTCA	GATCATTGAA	CGACTCCATG	9000
GTCTTAGCGC	ATTTACACTC	CACAGTTACT	CTCCAGGTGA	GATCAATAGG	9050
GTGGCTTCAT	GCCTCAGGAA	ACTTGGGGTA	CCACCTTTGC	GAACCTGGAG	9100
ACATCGGGCC	AGAAGTGTCC	GCGCTAAGCT	ACTGTCCCAG	GGGGGGAGGG	9150
CCGCCACTTG	TGGCAGATAC	CTCTTTAACT	GGGCAGTAAG	GACCAAGCCT	9200
AAACTCACTC	CAATCCCGGC	CGCGTCCCAG	CTGGACTTGT	CTGGCTGGTT	9250
CGTCGCTGGT	TACAGCGGGG	GAGACATATA	TCACAGCCTG	TCTCGTGCCC	9300
GACCCCGCTG	GTTCCTCGTG	TGCTTACTCC	TACTTTCTGT	AGGGGTAGGC	9350
ATTTACCTGC	TCCCCAACCG	ATGAACGGGG	AGCTAACCAC	TCCAGGCCTT	9400
AAGCCATTTC	CTGTTTTTTT	TTTTTTTTTT	TTTTTTTTTT	TCTTTTTTTT	9450
TTCTTTCTCT	TTCTTTCTTT	TTTTCCTTTC	TTTTTCCCTT	CTTTAATGGT	9500

FIG. 4E

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GGCTCCATCT	TAGCCCTAGT	CACGGCTAGC	TGTGAAAGGT	CGTGAGCCG	9550
CATGACTGCA	GAGAGTGCTG	ATACTGGCCT	CTCTGCAGAT	CATGT	9595

FIG. 4F

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RGSRPSWGFT	DPRRRSRNLG	KVIDTLTQGF	ADLMGYIPLV	GAPLGGAARA	150
LAHGVRVLED	GVNYATGNLP	GCSFSIFLLA	LLSCLTIPAS	AYEVRNVSGI	200
YHVINDCSNS	STVYEADVI	MHTPGCVPCV	QEGNSSROW	ALITPTLAARN	250
ASVPTTTIRR	HVDLLVGTA	FCSAMYVGL	CGSIFLVSQL	FTFSPPRHET	300
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GCPERMASCR	PIDWFAQGWG	PTTYTKFNSS	DQRPYCWHYA	PRPGVVPAS	500
QVCGPVYCF	PSPVVGITD	RSGVPTYSWG	ENETDMLLN	NIRPPQGNWF	550
GCTWMNSTGF	TKTCGGPPCN	IGGVGNRILI	CPIDCFRKHP	EATYTKCGSG	600
PWLTPRCLVD	YPYRLWHYPC	TLNFSIFKVR	MYVGGVEHRL	NAAONWIRGE	650
RONLEDRDRS	ELSPLLLSTT	EWQILPCAFT	TLPALSTGLI	HLHQNVVDVQ	700
YLYGVGSFAV	SFAIKWEYTL	LLFLLLADAR	VCACIWMMLL	IAQAEAALEN	750
LVVLNAASVA	GAHGILSFLV	FFCAAWYIKG	RLAPGAAYAF	YGVWPLLLLL	800
LALPPRAYAL	DREMAASCGG	AVLVGLVFLT	LSPYYKVFLT	RLIWWLQYFI	850
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FIG. 4G

SUBSTITUTE SHEET (RULE26)

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TCSNIWHGTF	PINAYTTGPC	TPSPAPNYSR	ALWRVAAEEY	VEVIRVGDFH	2100
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FIG. 4H

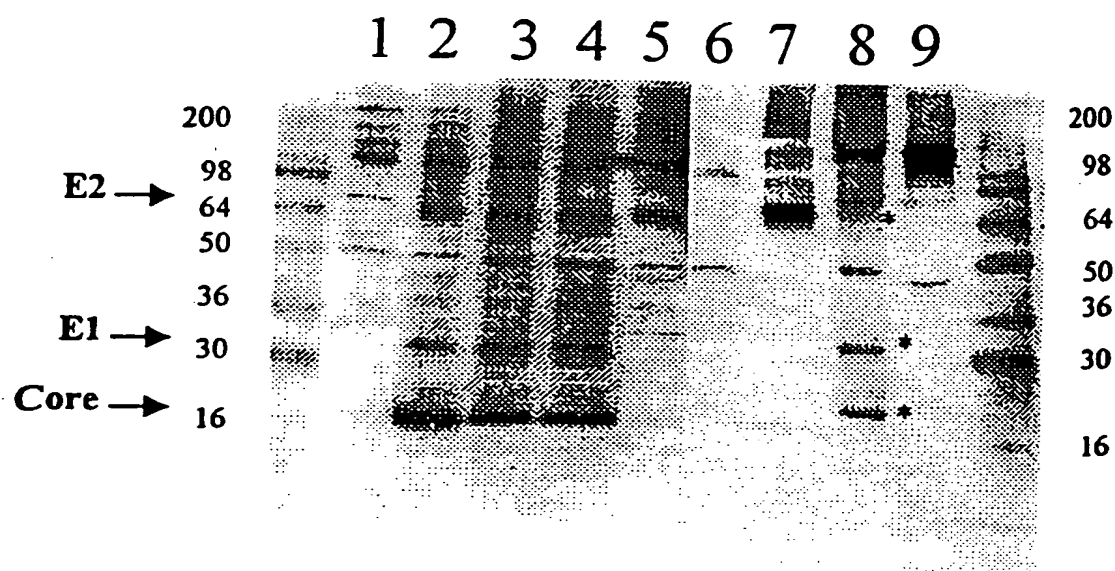


FIG. 5

SEQUENCE LISTING

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Bukh, Jens
Emerson, Suzanne
Purcell, Robert

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Ile Tyr Leu Lys Pro Gly Pro Leu Phe Tyr Gln Asp Tyr Lys Gly Pro
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Val Tyr His Arg Ala Pro Leu Glu Leu Phe Glu Glu Gly Ser Met Cys
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Glu Thr Thr Lys Arg Ile Gly Arg Val Thr Gly Ser Asp Gly Lys Leu
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Tyr His Ile Tyr Val Cys Ile Asp Gly Cys Ile Ile Ile Lys Ser Ala
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Lys Thr Lys Arg Asn Thr Asn Arg Arg Pro Gln Asp Val Lys Phe Pro
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Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ala Val Asp Ser Thr
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Val Arg Leu Thr Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr
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Thr Pro His Pro Asn Ile Glu Glu Val Ala Leu Gly Gln Glu Gly Glu
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Ile Pro Phe Tyr Gly Arg Ala Ile Pro Leu Ser Tyr Ile Lys Gly Gly
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Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala
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Asp Cys Asn Val Ala Val Thr Gln Val Val Asp Phe Ser Leu Asp Pro
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Thr Phe Thr Ile Thr Thr Gln Ile Val Pro Gln Asp Ala Val Ser Arg
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Ser Glu Thr Thr Val Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu
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Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala
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1795 1800 1805

Thr Thr Ile Leu Leu Asn Ile Leu Gly Gly Trp Leu Ala Ser Gln Ile
1810 1815 1820

Ala Pro Pro Ala Gly Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly
1825 1830 1835 1840

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1845 1850 1855

Ala Gly Tyr Gly Ala Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile
1860 1865 1870

Met Ser Gly Glu Lys Pro Ser Met Glu Asp Val Val Asn Leu Leu Pro
1875 1880 1885

Gly Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala
1890 1895 1900

Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met
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1925 1930 1935

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1940 1945 1950

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2705 2710 2715 2720

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Ala Arg Pro Arg Leu Leu Leu Phe Gly Leu Leu Leu Leu Phe Val Gly
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Val Gly Leu Phe Leu Leu Pro Ala Arg
3025 3030

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14 December 2000 (14.12.2000)

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7/01, C07K 14/18

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(30) Priority Data:
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Filed on **4 June 1999 (04.06.1999)**

(71) Applicant (for all designated States except US): **THE GOVERNMENT OF THE UNITED STATES OF AMERICA** as represented by **THE SECRETARY, DEPARTMENT OF HEALTH AND HUMAN SERVICES [US/US]**; Office of Technology Transfer, National Institutes of Health, Suite 325, 6011 Executive Boulevard, Rockville, MD 20852 (US).

(72) Inventors; and

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(74) Agents: **FEILER, William, S. et al.**; Morgan & Finnegan, L.L.P., 345 Park Avenue, New York, NY 10154 (US).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

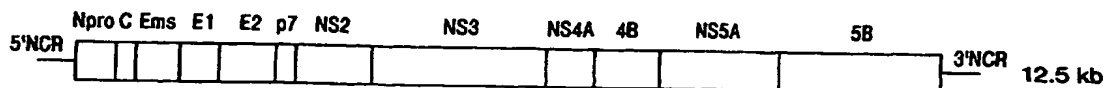
— with international search report

(88) Date of publication of the international search report:
15 November 2001

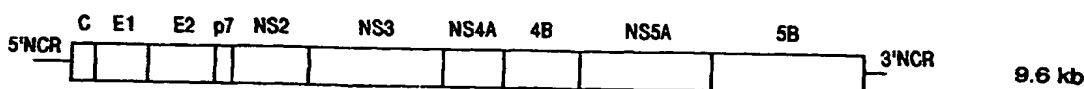
[Continued on next page]

(54) Title: **HCV/BVDV CHIMERIC GENOMES AND USES THEREOF**

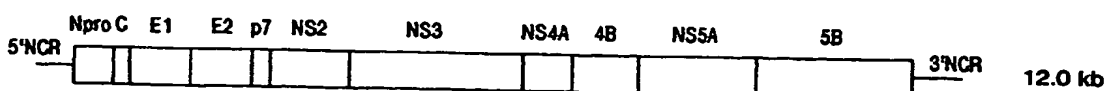
BVDV-NADL



HCV-H77C



HCV/BVDV (Chimeric RNA)



(57) Abstract: The present invention relates to molecular approaches to the production of nucleic acid sequences which comprise the genomes of chimeric hepatitis C virus-bovine viral diarrhea viruses (HCV-BVDV). The invention also relates to the use of these chimeric nucleic acid sequences to produce chimeric virions in cells and the use of these chimeric virions in HCV antibody neutralization assays, and for the development of vaccines and therapeutics for HCV.

WO 00/75352 A3



For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/15527

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/86 C12N7/01 C07K14/18

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, BIOSIS, EPO-Internal

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>FROLOV I ET AL: "CIS-ACTING RNA ELEMENTS REQUIRED FOR REPLICATION OF BOVINE VIRAL DIARRHEA VIRUS-HEPATITIS C VIRUS 5' NONTRANSLATED REGION CHIMERAS" RNA, CAMBRIDGE UNIVERSITY PRESS, CAMBRIDGE, GB, vol. 4, no. 11, 25 November 1998 (1998-11-25), pages 1418-1435, XP000952790 ISSN: 1355-8382 the whole document</p> <p style="text-align: center;">--- -/--</p>	1, 7, 8, 11, 12



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents :

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *G* document member of the same patent family

Date of the actual completion of the international search

5 February 2001

Date of mailing of the international search report

14. 02. 2001

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Chambonnet, F

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/15527

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>H LU ET AL: "Poliovirus chimeras replicating under the translation control of genetic elements of HCV reveal unusual properties of the IRES of HCV" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 93, February 1996 (1996-02), pages 1412-1417, XP002919370 ISSN: 0027-8424 the whole document</p>	12,13
X	<p>VENUGOPAL K. & GOULD E.A.: "Towards a new generation of Flavivirus vaccines" VACCINE, vol. 2, no. 11, 1994, pages 966-975, XP002919372 GUILDFORD GB the whole document</p>	11,12,20
P, X	<p>WO 99 55366 A (FROLOV ILYA ;MCBRIDE M SCOTT (US); RICE CHARLES M (US); UNIV WASHI) 4 November 1999 (1999-11-04) page 4, line 21 - line 30 page 10, line 31 -page 11, line 17 page 11, line 33 -page 15, line 8; claims 1-10,16-21; figures 21,25,26; examples 1,2,4,5</p>	1,2,7,8, 14-21
A	<p>MEYERS G ET AL: "RECOVERY OF CYTOPATHOGENIC AND NONCYTOPATHOGENIC BOVINE VIRAL DIARRHEA VIRUSES FROM CDNA CONSTRUCTS" JOURNAL OF VIROLOGY, THE AMERICAN SOCIETY FOR MICROBIOLOGY, US, vol. 70, no. 12, December 1996 (1996-12), pages 8606-8613, XP000952807 ISSN: 0022-538X the whole document</p>	7
A	<p>YU H ET AL: "SEQUENCE AND STRUCTURAL ELEMENTS AT THE 3' TERMINUS OF BOVINE VIRALDIARRHEA VIRUS GENOMIC RNA: FUNCTIONAL ROLE DURING RNA REPLICATION" JOURNAL OF VIROLOGY, THE AMERICAN SOCIETY FOR MICROBIOLOGY, US, vol. 73, no. 5, May 1999 (1999-05), pages 3638-3648, XP000946998 ISSN: 0022-538X the whole document</p>	7

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INTERNATIONAL SEARCH REPORT

Int. Patent Application No

PCT/US 00/15527

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
T	<p>LAI VC, ZHONG W, SKELTON A, INGRAVALLO P, VASSILEV V, DONIS RO, HONG Z, LAU JY: "Generation and characterization of a hepatitis C virus NS3 protease-dependent bovine viral diarrhea virus." JOURNAL OF VIROLOGY., vol. 74, no. 14, July 2000 (2000-07), pages 6339-6347, XP000952808 THE AMERICAN SOCIETY FOR MICROBIOLOGY., US ISSN: 0022-538X the whole document</p> <p>-----</p>	7,8

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 00/15527

B x I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

B x II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☒ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-6 11-13 partially 9, 10, 14-21

A nucleic acid molecule comprising a chimeric virus genome, said genome being a BVDV genome in which the structural region of the BVDV genome has been replaced by the structural region of a hepatitis C virus genome; a DNA construct comprising said molecule; an RNA transcript of said DNA construct; a host cell transfected with said DNA construct or RNA transcript; a chimeric HCV-BVDV produced by said host cell; a composition comprising said virus.

2. Claims: 7, 8 and partially 9, 10, 14-21

A nucleic acid molecule comprising a chimeric virus genome, said genome being a BVDV genome in which the non-structural region of the BVDV genome has been replaced by the non-structural region of a hepatitis C virus genome; a DNA construct comprising said molecule; an RNA transcript of said DNA construct; a host cell transfected with said DNA construct or RNA transcript; a chimeric HCV-BVDV produced by said host cell; a composition comprising said virus.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 00/15527

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9955366 A	04-11-1999	AU 3757199 A EP 1071454 A	16-11-1999 31-01-2001